

BIO-212 - Lecture 3

Introduction to Nucleic Acids

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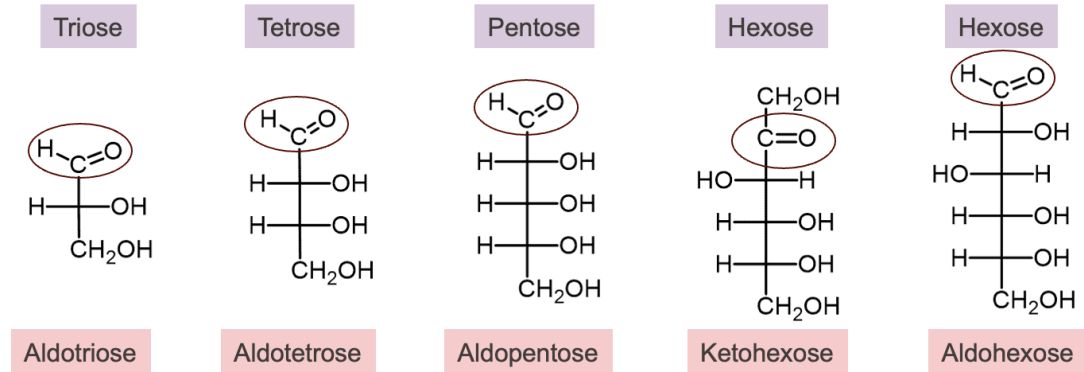
Laboratory of Virology and Structural Immunology
Global Health Institute, School of Life Sciences
École Polytechnique Fédérale de Lausanne



Lecture 2 – Quick Summary

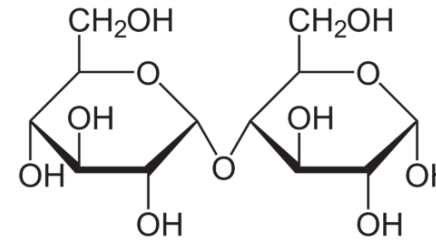
• Carbohydrate building blocks

Monosaccharides:

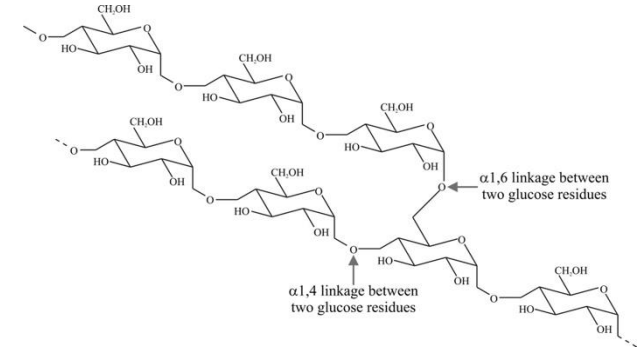


• Carbohydrate polymers and their roles

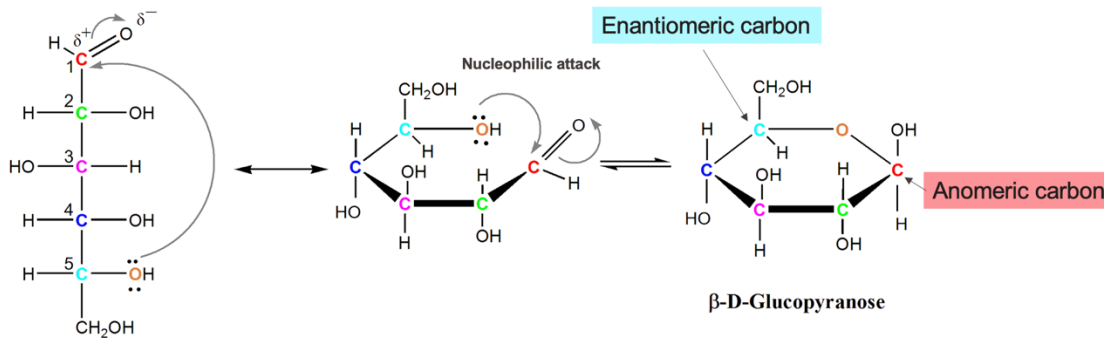
Disaccharide (Maltose)



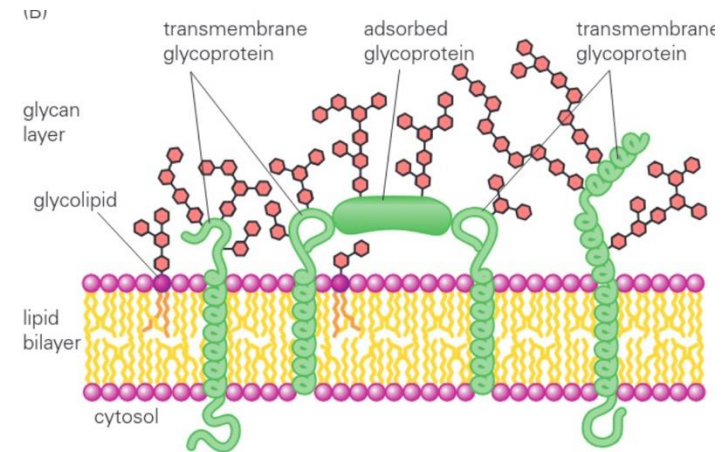
Polysaccharide (Amylopectin)



Hemiacetal and hemiketal forms in solution:

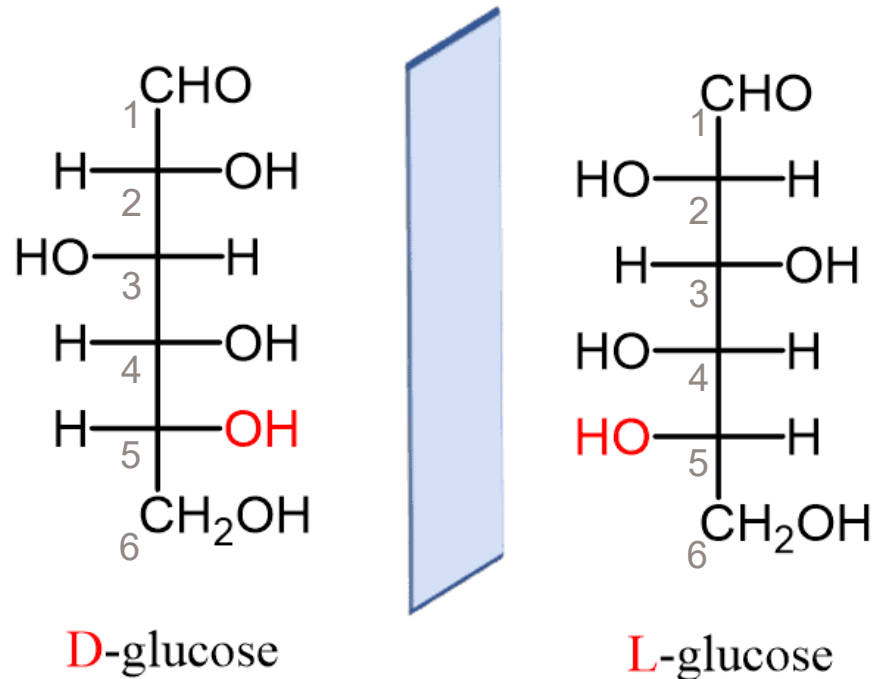


Glycolipids and glycoproteins



Lecture 2 - Extra Clarifications

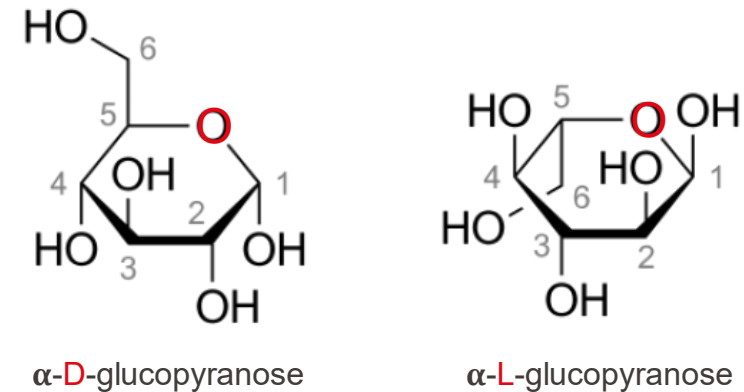
• Enantiomeric forms of carbohydrates



- Enantiomers of monosaccharides are connected by mirror symmetry at every chiral center.
- The main chiral center (C5 in glucose) is just used to assign D- or L- form

• The position of terminal -CH₂OH

- In the cyclic form the position of terminal -CH₂OH group will depend on the D- or L- isomerism:

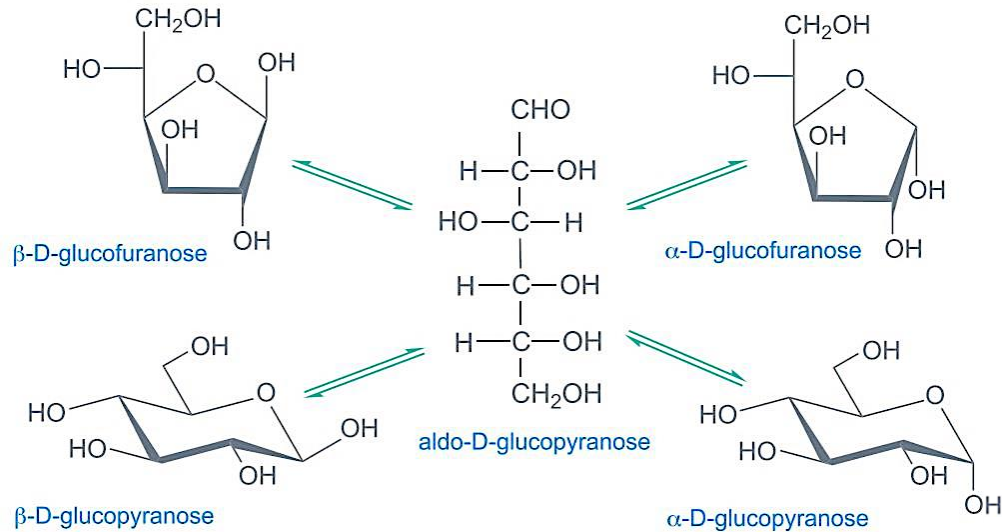


- This is counter-intuitive given the orientation of H at position C5 (CH₂OH expected to face down in D-form and up in L-form, but that does not happen)
- The final orientation is influenced by the rotation around the C4-C5 bond during cycle assembly.
- See: <https://www.youtube.com/watch?v=kXnkjLAvGwk>

Lecture 2 - Extra Clarifications

Multiple carbon options for cyclic state assembly

- In theory any OH group at or after the 5th carbon can be used for assembly into cyclic form

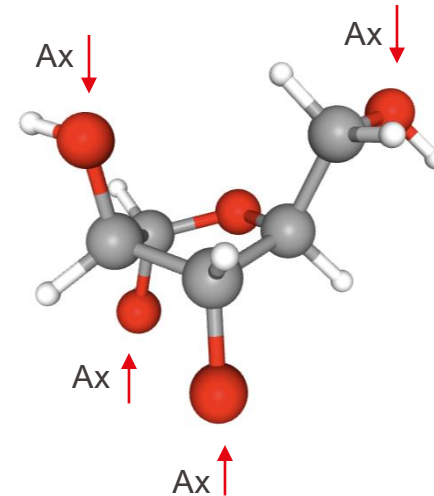


- In the majority of cases the most stable linkage is via **pen-ultimate carbon** (e.g., C5 in glucose)
- 5- and 6-membered rings are energetically favored but preference for one or the other depends on the specific monosaccharide and orientation of its OH groups

Achieving maximum distance of electron-rich groups

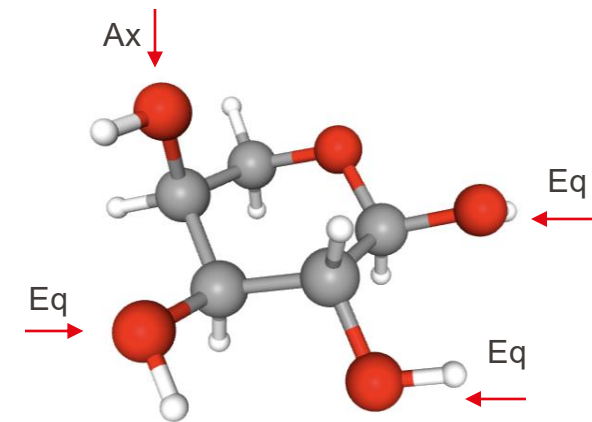
- State of minimum energy is the one with maximum spacing between electron-rich groups
- Example: **L-arabinose** (an aldopentose)

Alpha-L-Arabinofuranose



10-20% of molecules in solution
(combined alpha and beta states)

Alpha-L-Arabinopyranose



80-90% of molecules in solution
(combined alpha and beta states)

- Equatorial orientation (right) provides better spacing between OH groups compared to axial (left)

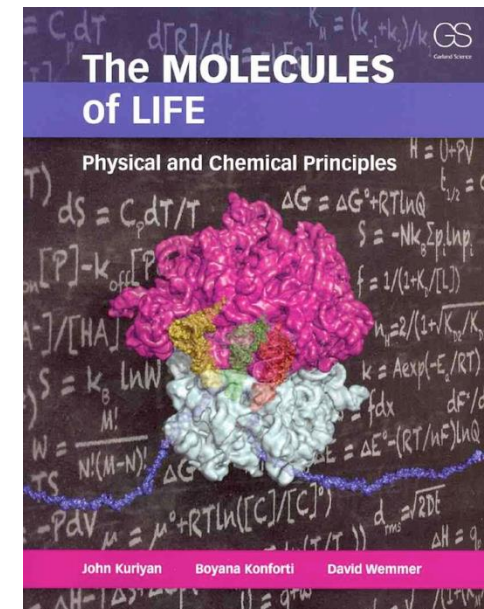
Lecture 3 - Outline

Today:

- Nucleotide building blocks
- Assembly into nucleic acids
- Relevant covalent and non-covalent interactions

Reading suggestions:

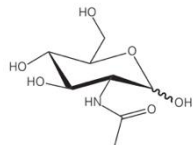
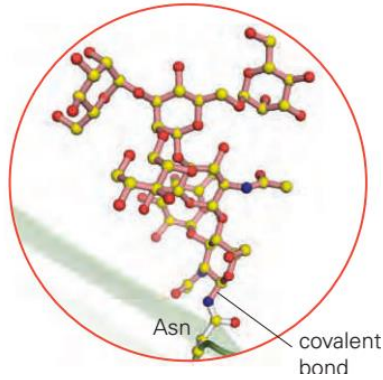
- The Molecules of Life (Chapter 1, 19)



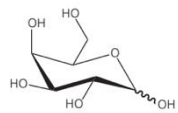
The molecules of Life

Macromolecular Structure

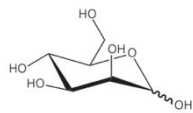
Carbohydrates



N-acetylglucosamine (GlcNAc)



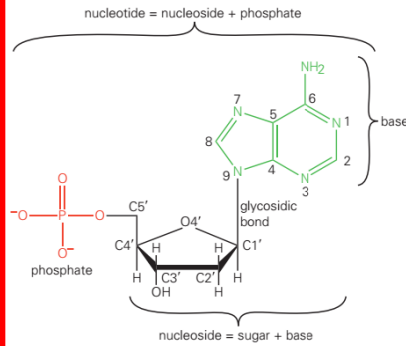
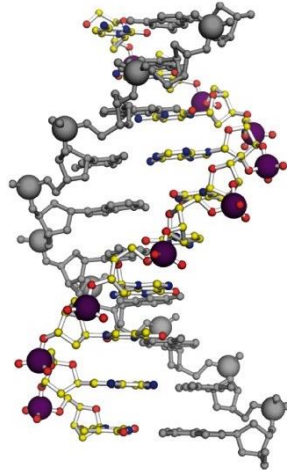
galactose (Gal)



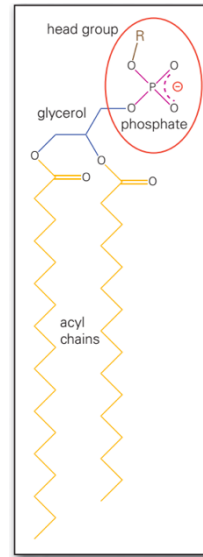
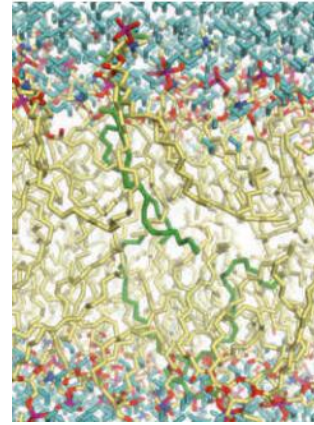
mannose (Man)

Building Block

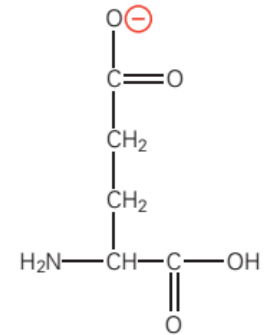
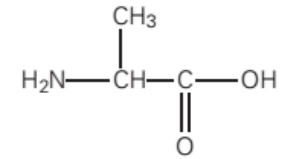
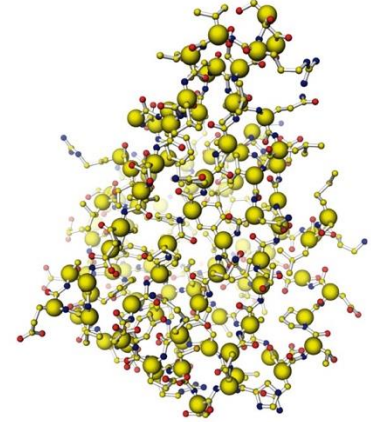
Nucleic Acids



Lipids

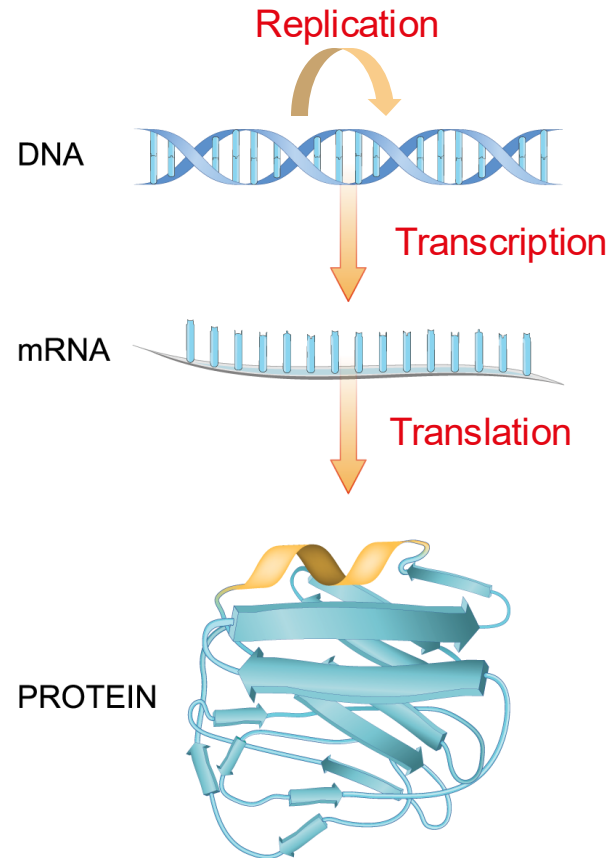
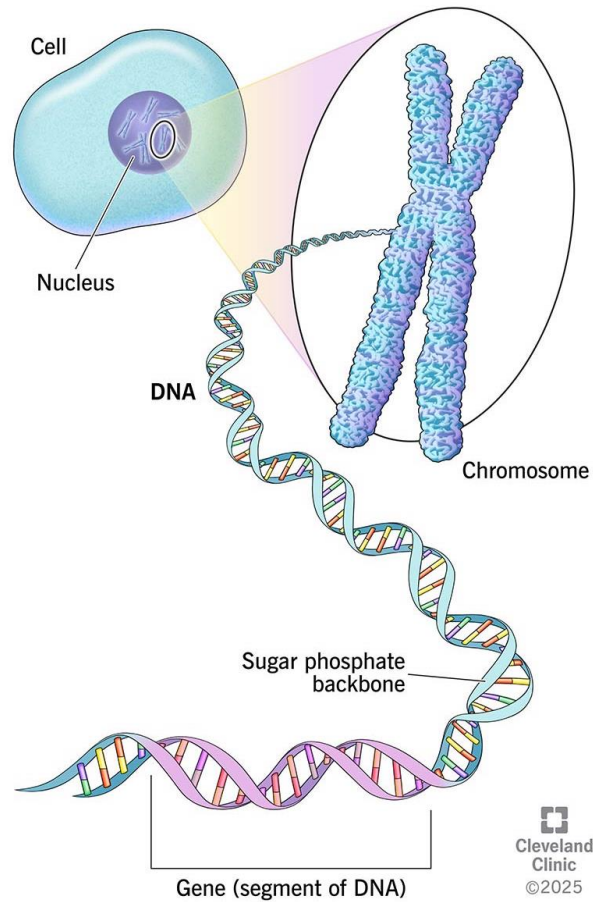


Proteins



Nucleic acids in cells

- The main biological functions are **storage**, **transmission** and **expression** of genetic information



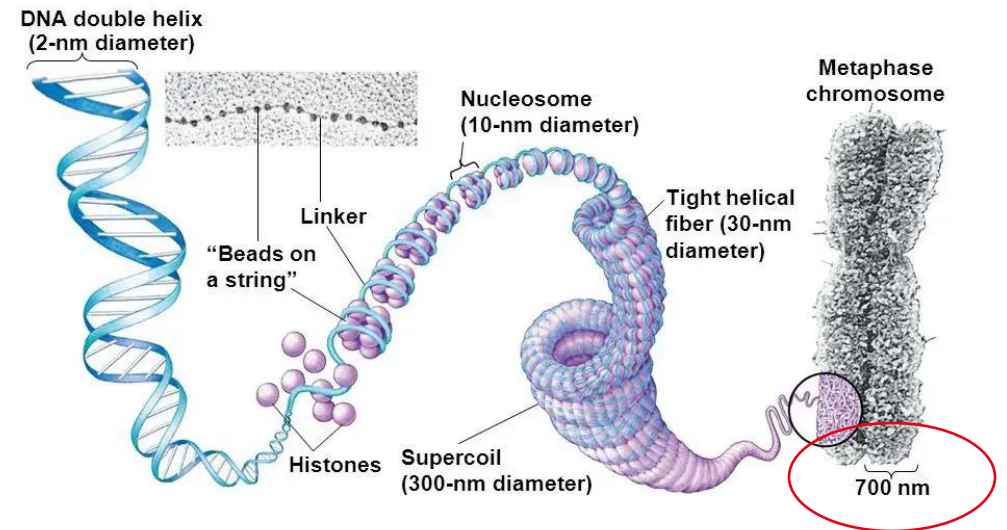
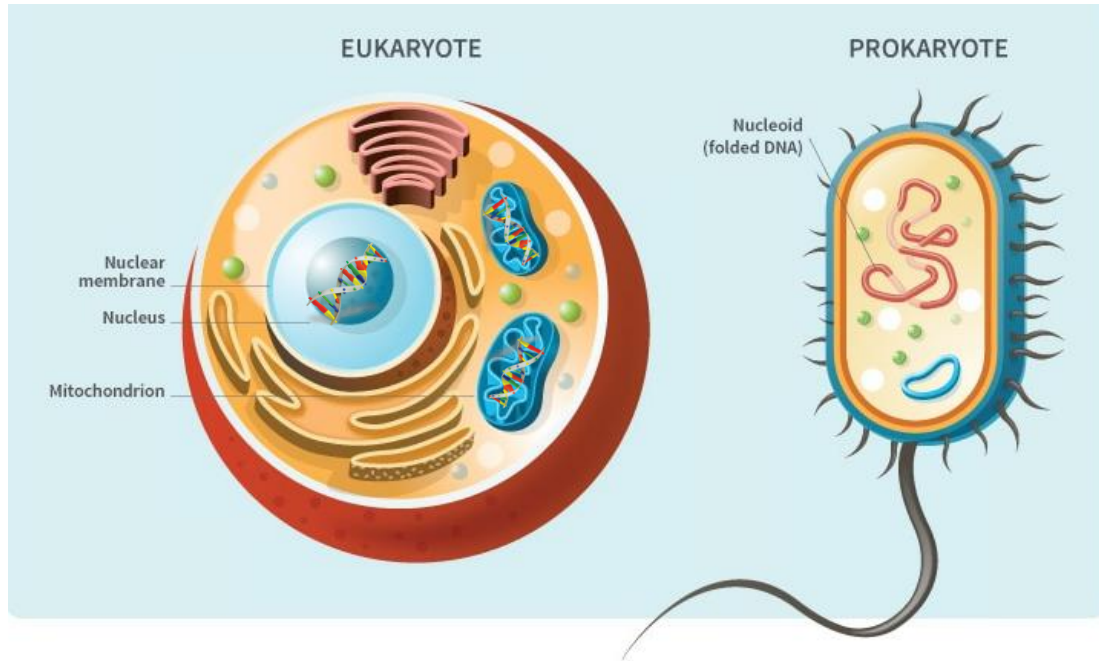
Johannes Friedrich Miescher
(1844-1895)

First person to isolate nucleic acids and suggest a role in heredity (“nuclein”)

- Nucleotides also serve as **energy carriers** and **signaling molecules**, while RNA has many **regulatory**, **catalytic**, and **sensing** roles (particularly important for translation).

Nucleic acids in cells

- In eukaryotes most **DNA** is stored as chromatin within the nuclear envelope, and in organelles such as mitochondria and chloroplasts (plants/algae)
- **RNA** exists in the nucleus, cytosol and inside certain organelles (e.g., mitochondria, exosomes, chloroplasts etc.)



The length of human DNA in extended state = 2.2 m
Diameter of the cell nucleus: 5-10 μm

- Human haploid genome is composed of 3.2×10^9 nucleotide pairs arranged into $\sim 10^7$ nucleosomes which assemble into 23 chromosomes
- Helical organization and packaging with histones reduces the effective size by 5-6 orders of mag.

Nucleotides – The building blocks of nucleic acids

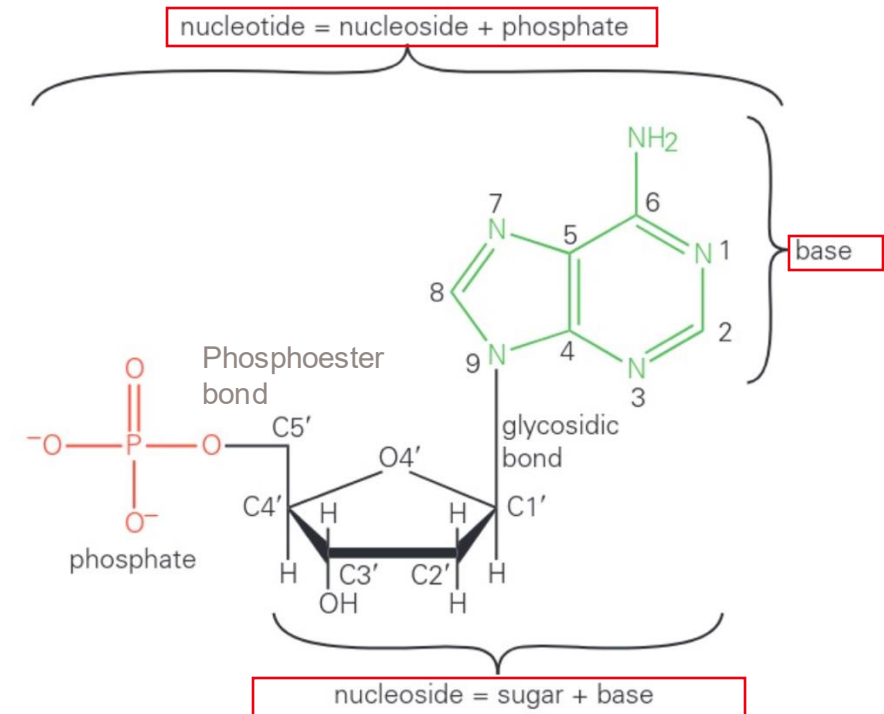
- DNA and RNA are both polymers of **nucleotides**.
- They can be synthesized endogenously and are therefore non-essential nutrients

Linkage between the base and carbohydrate:

- **N-glycosidic** (similar to O-glycosidic but via Nitrogen)

Linkage between the phosphate and carbohydrate:

- **Phosphoester** (hydroxyl group to phosphate)

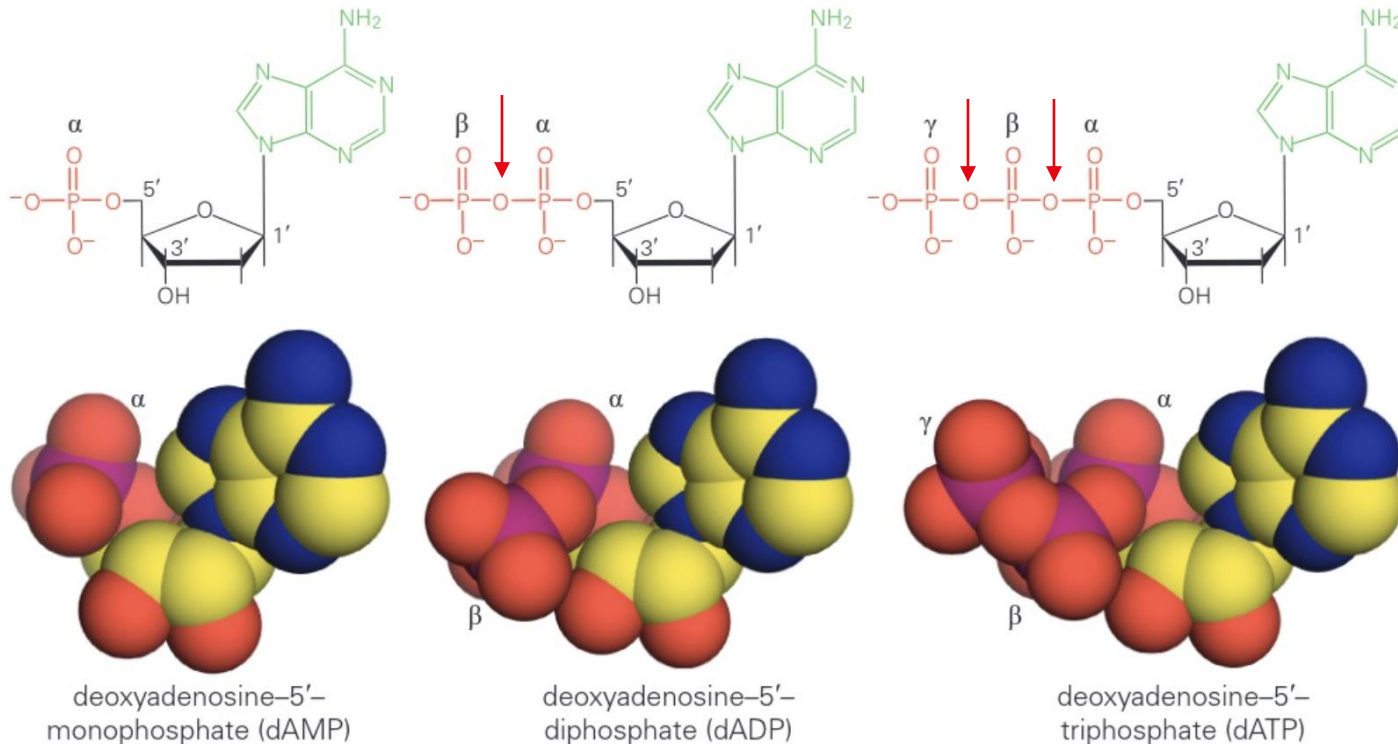
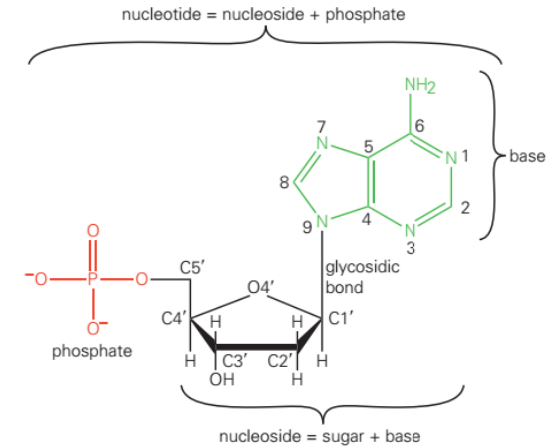


- Each nucleotide consists of the following functional groups:
 - 1) **Aldopentose** monosaccharide (black)
 - 2) Nitrogen-containing aromatic ring system, i.e. **Lewis base** (green)
 - 3) **Phosphate** group (red)

Note the difference between these chemical entities

Nucleotides – The phosphate group(s)

- Nucleotides with one, two or three phosphate groups are referred to as nucleotide **mono-**, **di-** or **tri-phosphate**.
- The three phosphate groups are called **alpha**, **beta** and **gamma**



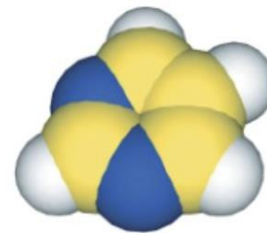
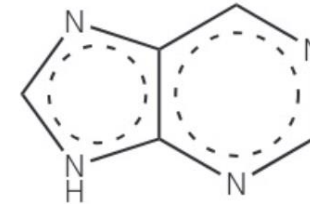
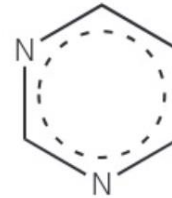
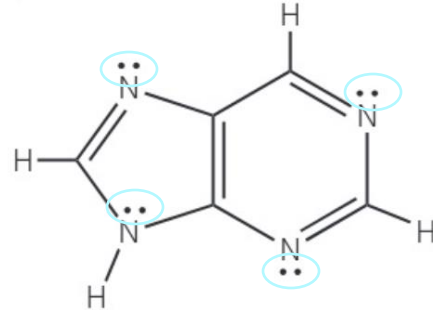
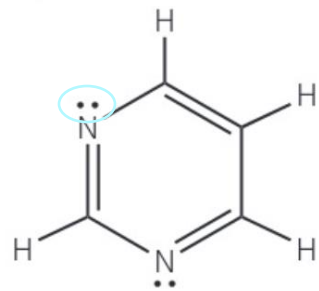
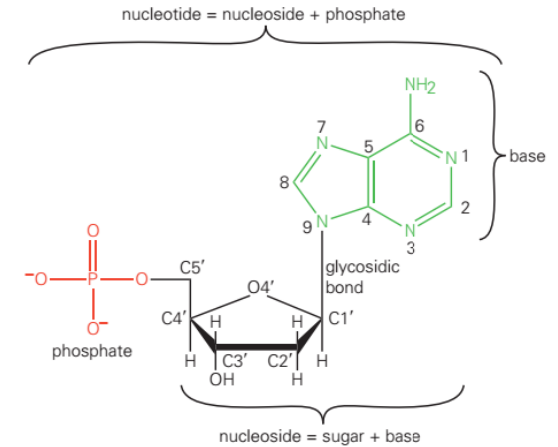
Phosphate groups are linked via **phosphoanhydride bonds**

Breakage of these bonds (i.e., in ATP) produces **~30-40 kJ/mol** of energy.

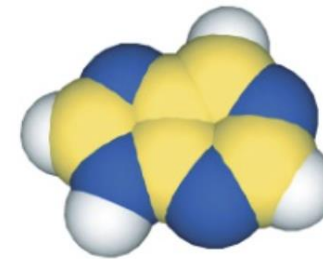
This is a fundamental process in **cell metabolism**.

Nucleotides – The Bases

- The nucleotides comprising DNA and RNA are built with **5 different bases** (4 per nucleic acid type)
- The name “**base**” comes from its chemical composition - The ring systems contain **lone pairs of electrons in nitrogen atoms** being able to act as electron pair donors – so called **Lewis bases**



pyrimidine



purine

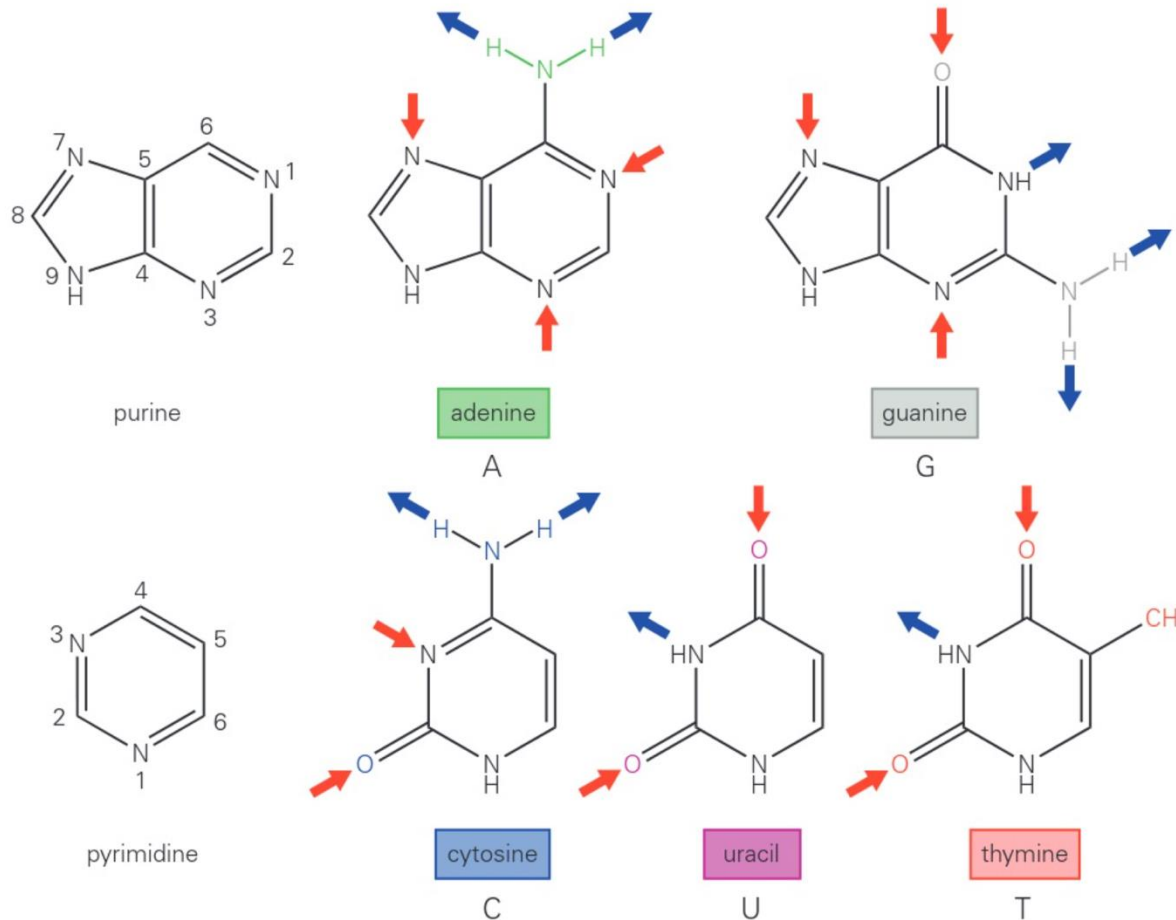
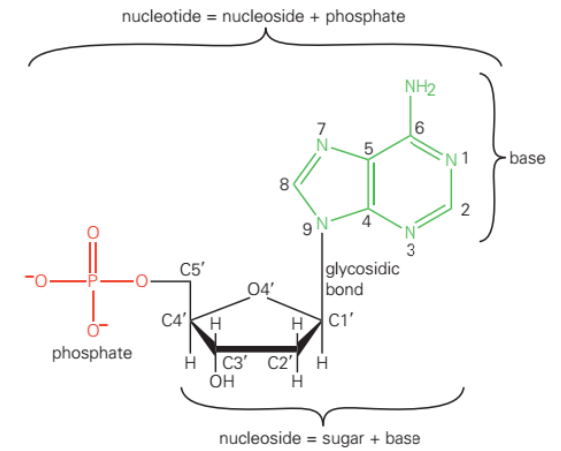
The rings are sp² hybridized making them planar.

Free electrons in N atoms make them potential hydrogen bond acceptors

- Nucleotide bases in RNA and DNA are substituted forms of two heterocyclic molecules known as **pyrimidine** and **purine**

Nucleotides – The Bases

- DNA contains two substituted purines (**adenine** and **guanine**)
- DNA contains two substituted pyrimidines (**cytosine** and **thymine**)
- In RNA **thymine** is replaced by **uracil**



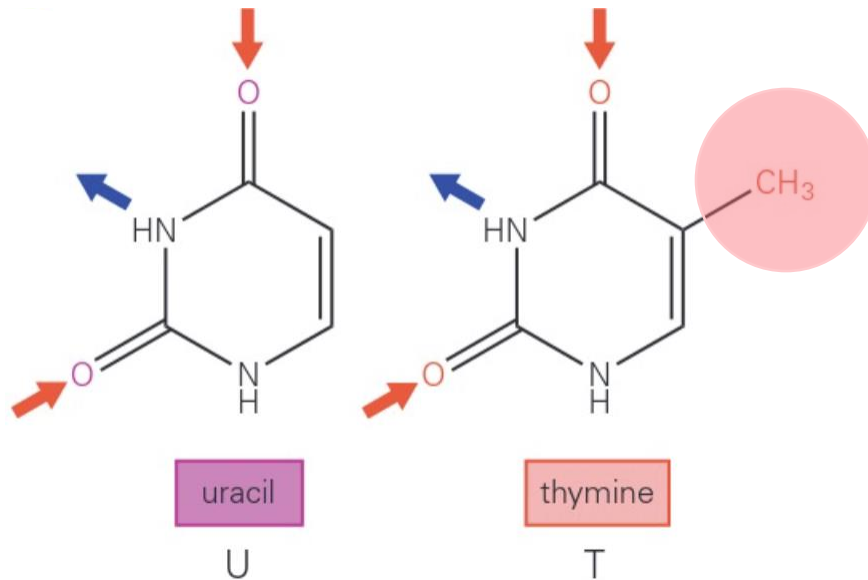
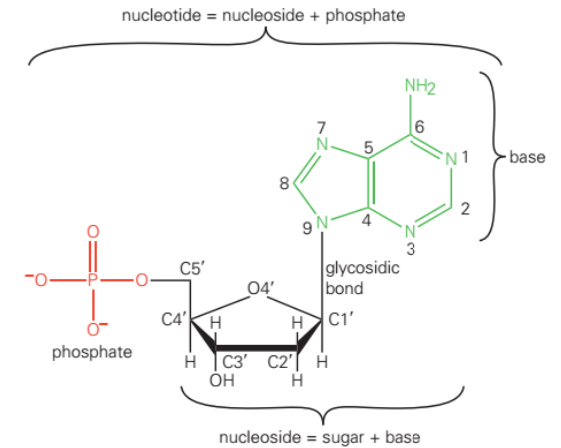
➔ Hydrogen bond acceptors

➔ Hydrogen bond donors

The unique distribution of hydrogen bond donors and acceptors in each base are key to DNA replication and transcription

Nucleotides – Thymine vs Uracil

- **Thymine** and **Uracil** feature the same hydrogen bond donor/acceptor organization, but thymine has one extra methyl (CH_3) group
- They replace each other in **DNA (T)** versus **RNA (U)** but are mutually exclusive



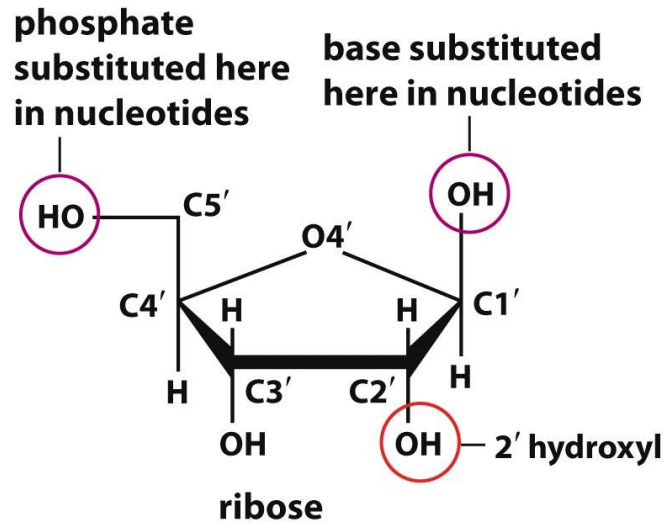
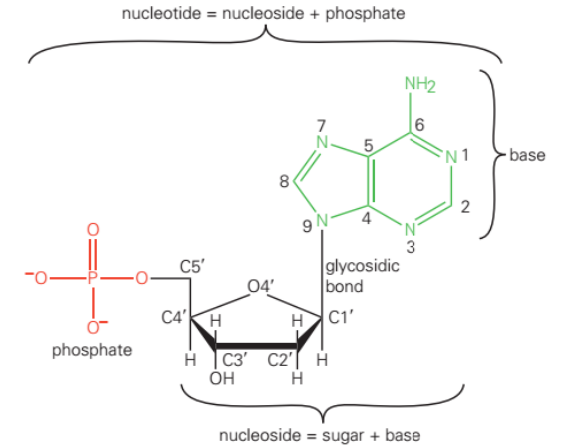
The extra **methyl group** present in thymine plays several important roles:

- Differentiation of DNA vs RNA by proteins
- Better packing of DNA helix due to hydrophobicity
- Detection of C→U conversion in DNA which reduces the mutation rate (improved genomic stability)
- RNA is meant to be short-lived intermediate so it uses “cheaper” uracil requiring fewer reactions to produce

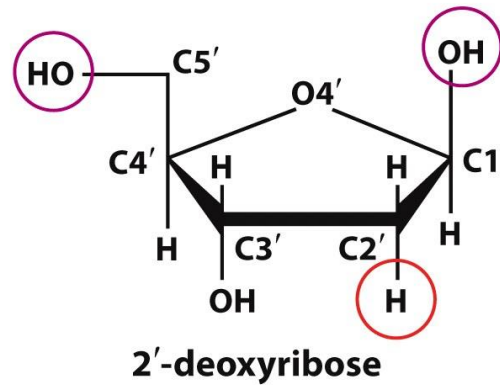
- Interestingly, **other bases (e.g., cytosine) can get methylated** by certain cellular enzymes and such modifications are the basis of **epigenetic** and **gene expression regulation**.

Nucleotides – The pentose sugar

- **Ribose** is the carbohydrate used in **RNA**.
- **2'-deoxy-ribose** is the carbohydrate used in **DNA**.



Ribonucleotides



Deoxyribonucleotides

The **2' hydroxyl (OH)** group in ribose causes several important differences:

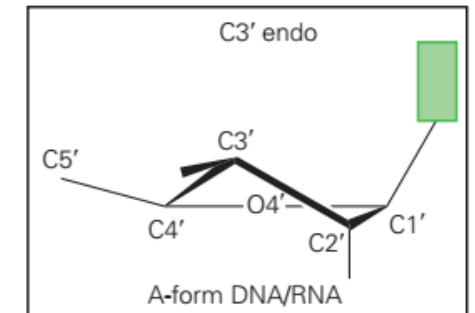
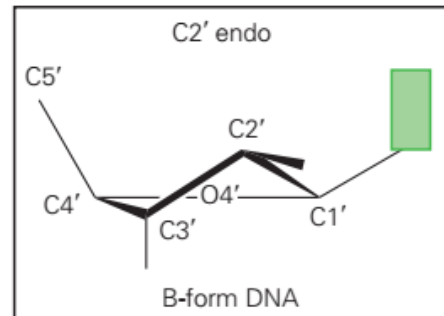
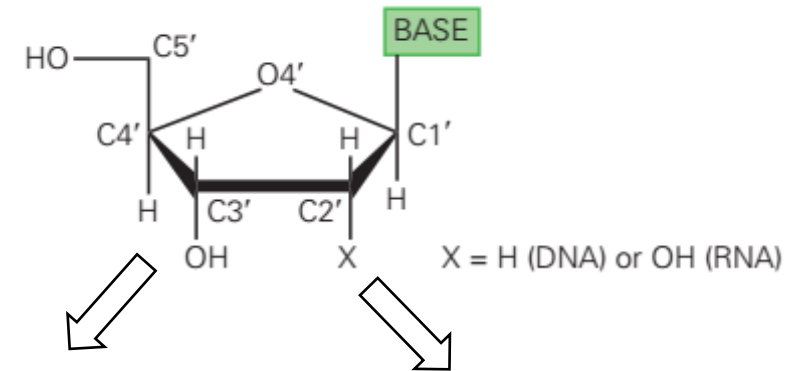
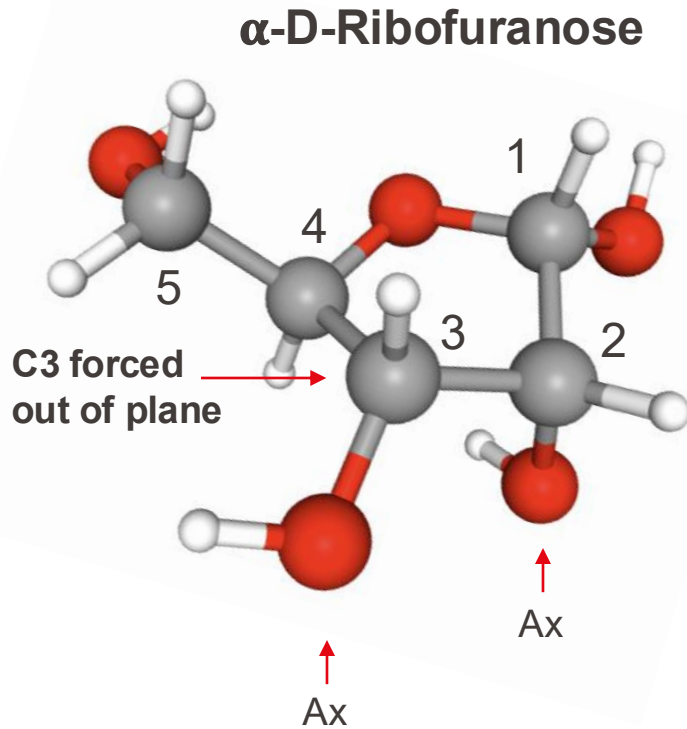
- OH can attack adjacent phosphate group in RNA causing spontaneous cleavage
- The repulsion with 3'OH causes forces C3 carbon out of the plane (C3'-pucker)
- Provides extra hydrogen bond donor and acceptor group useful for RNA recognition

RNA = Ribonucleic Acid.

DNA = Deoxyribonucleic Acid

Nucleotides – The pentose sugar

- **Furanose** sugars exist in the so-called “**envelope**” conformation which causes one group to be displaced out of the plane (“**pucker**”)
- In energetically favorable conformations four of the atoms of the pentose ring are roughly coplanar and one is out of the plane

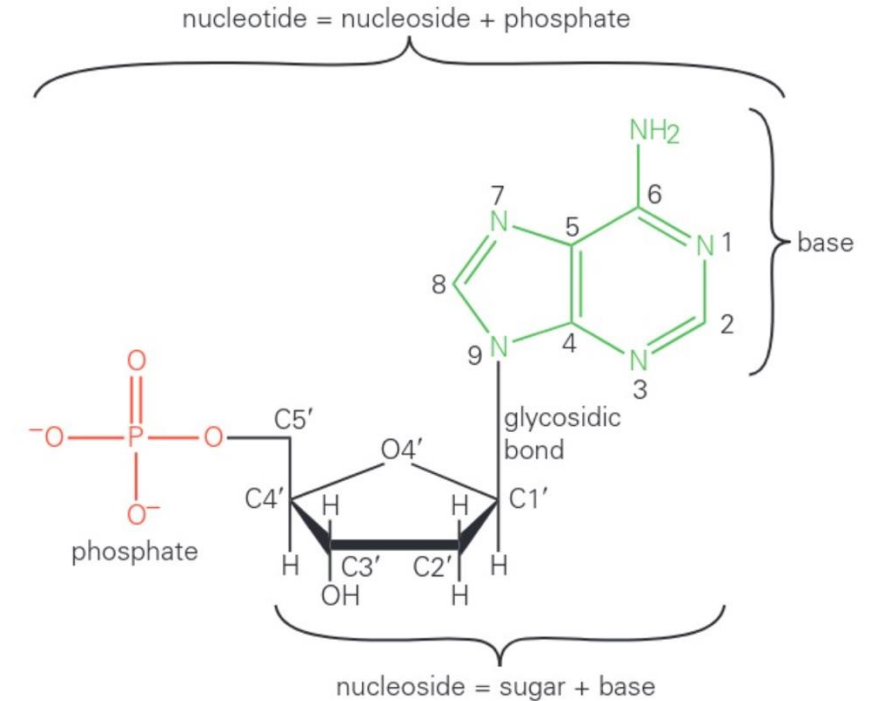


- In **DNA** and **RNA** molecules the pentose adopts a different sugar pucker conformation

Nucleotide naming

- Please note the name distinctions between **bases**, **nucleosides** and **nucleotides**

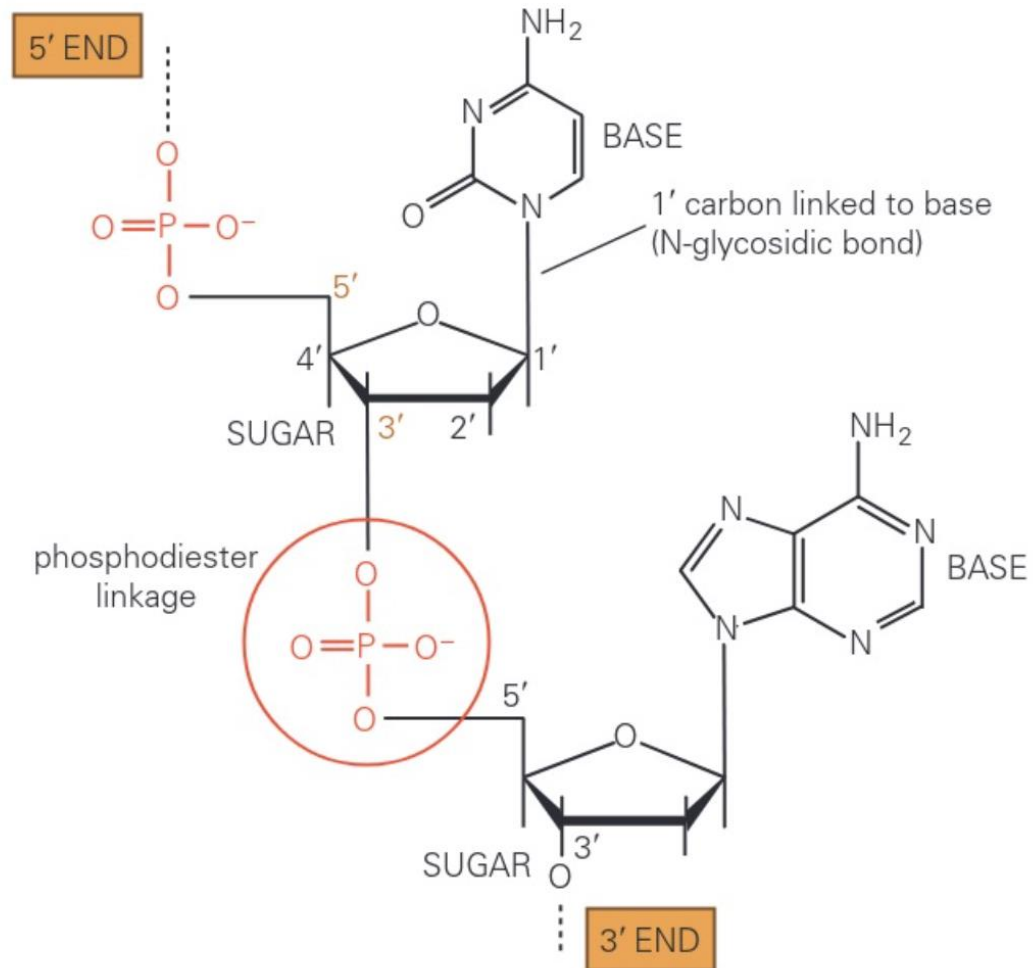
| Base | Nucleosides | Nucleotides |
|--------------|--------------------|--|
| RNA | | |
| Adenine (A) | Adenosine (A) | Adenosine 5'-monophosphate (AMP) |
| Guanine (G) | Guanosine (G) | Guanosine 5'-monophosphate (GMP) |
| Cytosine (C) | Cytidine (C) | Cytidine 5'-monophosphate (CMP) |
| Uracil (U) | Uridine (U) | Uridine 5'-monophosphate (UMP) |
| DNA | | |
| Adenine (A) | Deoxyadenosine (A) | Deoxyadenosine 5'-monophosphate (dAMP) |
| Guanine (G) | Deoxyguanosine (G) | Deoxyguanosine 5'-monophosphate (dGMP) |
| Cytosine (C) | Deoxycytidine (C) | Deoxycytidine 5'-monophosphate (dCMP) |
| Thymine (T) | Deoxythymidine (T) | Deoxythymidine 5'-monophosphate (dTMP) |



It is very important that you know the structures of all 5 bases, nucleotides and nucleosides!!!

Nucleic acids are polymers of nucleotides

- Nucleotides are joined together in DNA and RNA by the formation of a **phosphodiester bond** between the **3' carbon of one nucleotide and the 5' of the subsequent nucleotide**



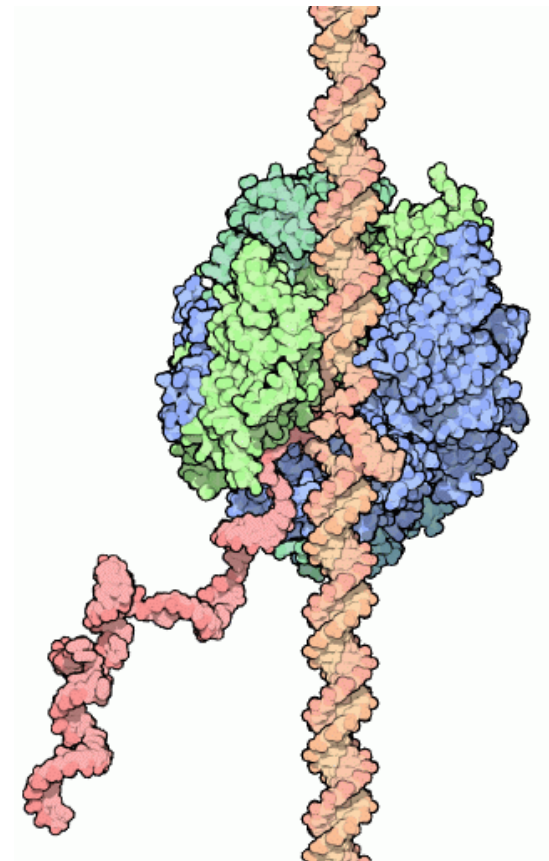
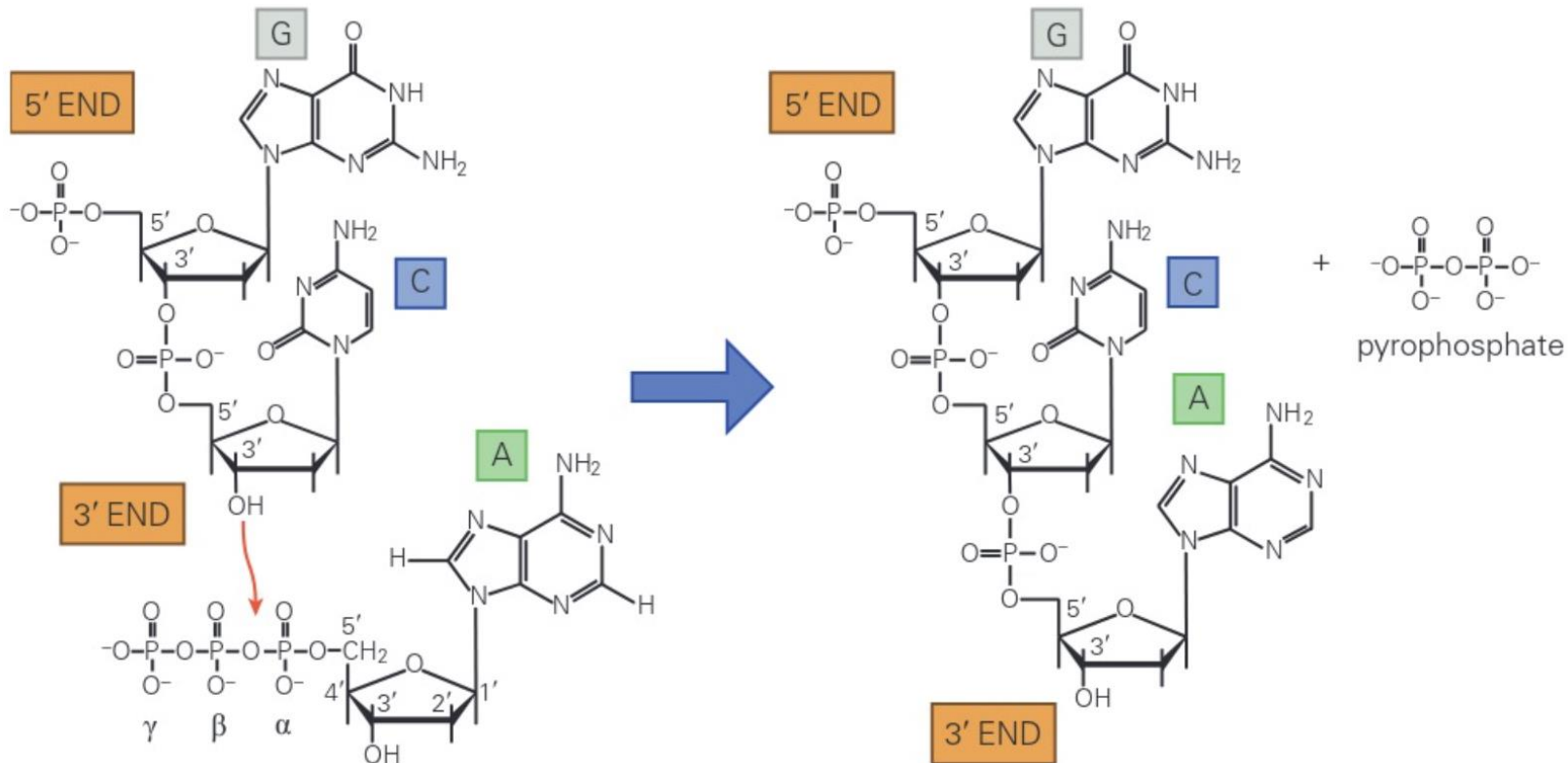
- The phosphate groups are **negatively charged** (anion nature) which represent an important determinant of the 3D structure of nucleic acids

- Phosphate-Sugar** linkages represent the backbone of nucleic acid polymers (repeating pattern of identical 2 units)

- The **bases remain accessible** with the potential hydrogen bond donors/acceptors being relatively unhindered by the backbone (rotation possible around glycosidic bond).

Stepwise addition of nucleotides

- The synthesis of new molecules of DNA and RNA involves the stepwise addition of nucleotides to one end of the chain. The enzymes that catalyze this reaction are called **polymerases**.
- The **triphosphate group** is high in energy and the hydrolysis of **phosphoanhydride bonds** drives the reaction

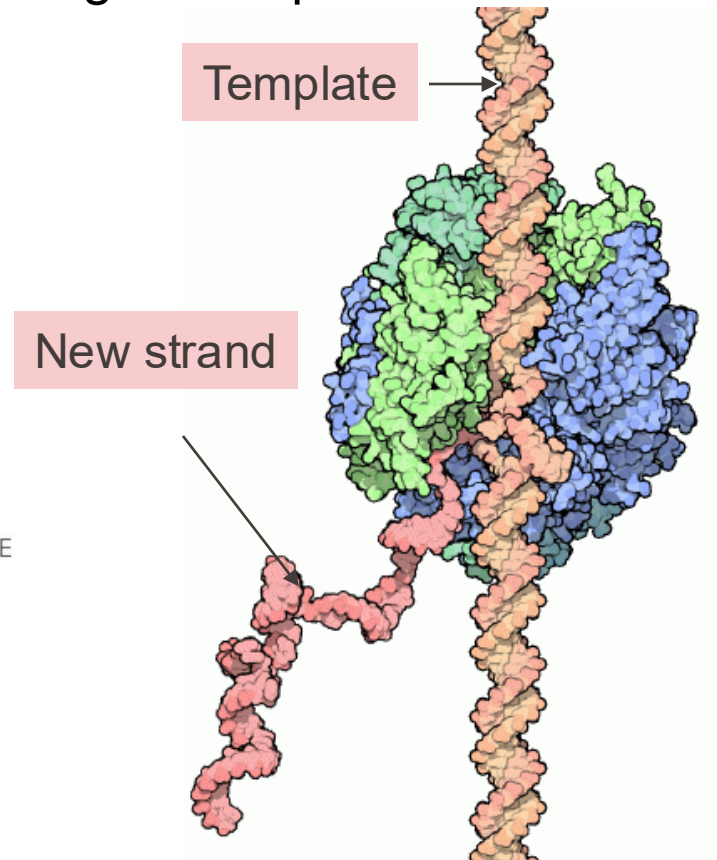
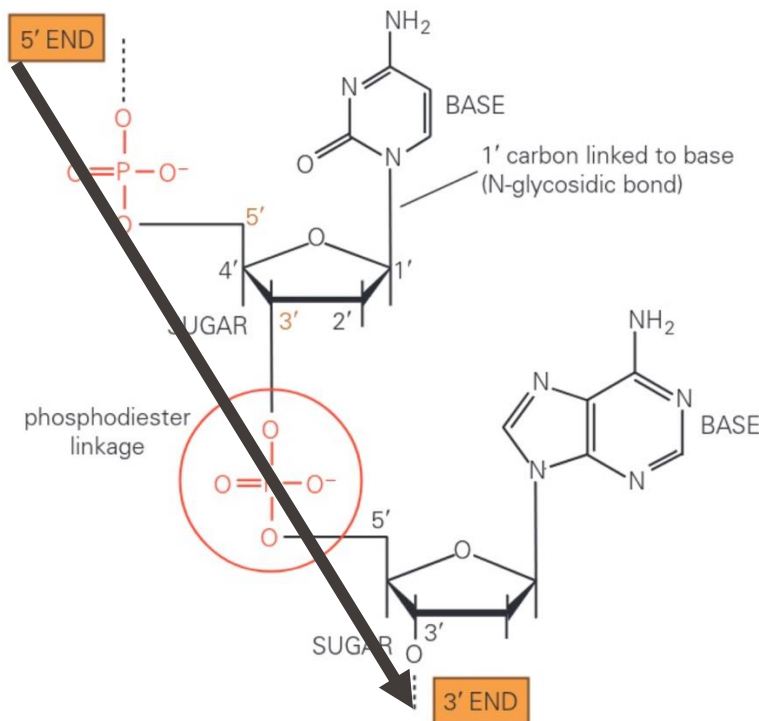


Model of RNA polymerase in action
(adapted from rcsb.org).

Stepwise addition of nucleotides

- DNA and RNA synthesis are **template directed** – The polymerases use a template strand to select each nucleotide to be added to the newly synthesized strand.
- This template-based assembly represents the essence of genetic data transfer during cell division as well as protein synthesis based on the corresponding DNA gene sequence

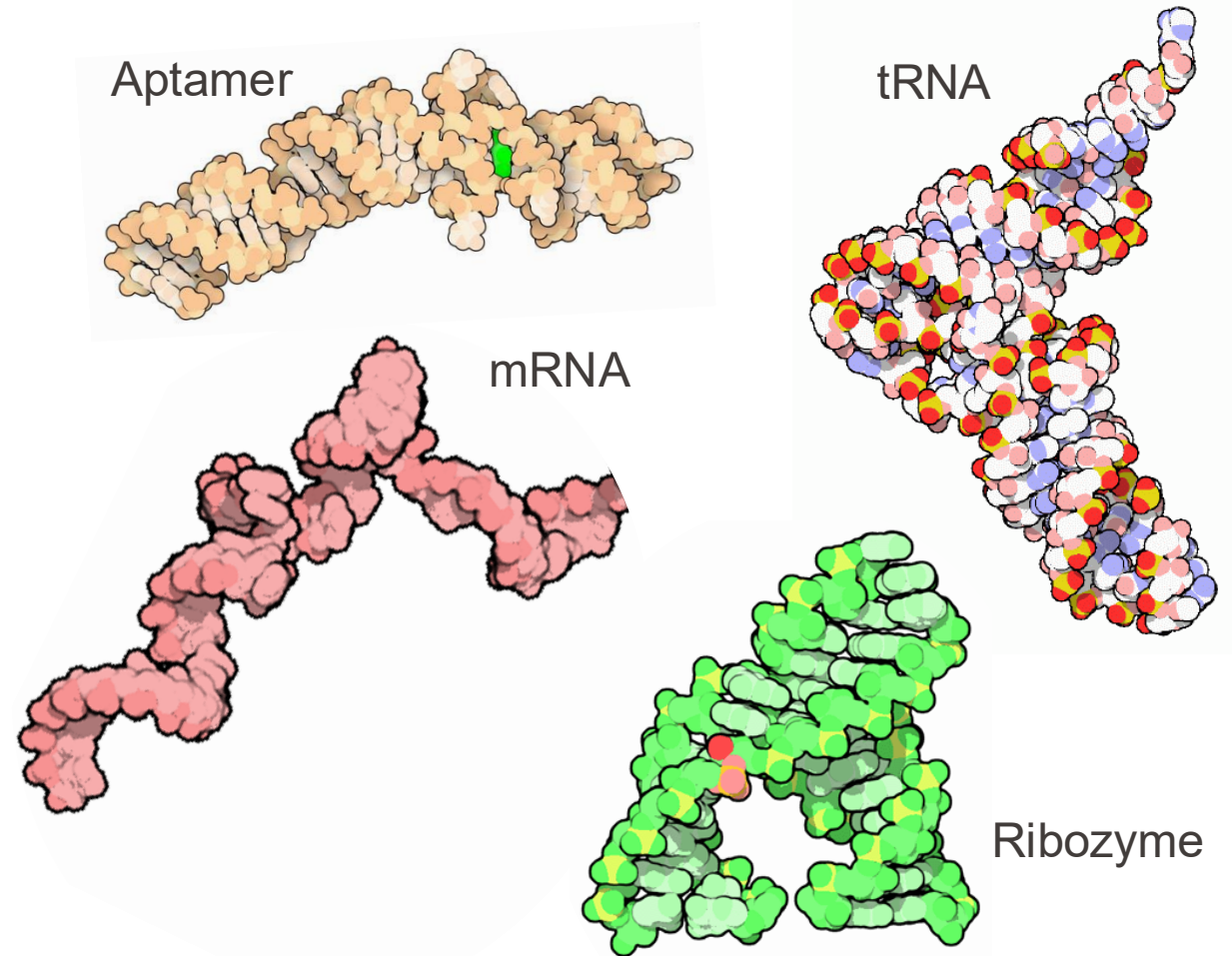
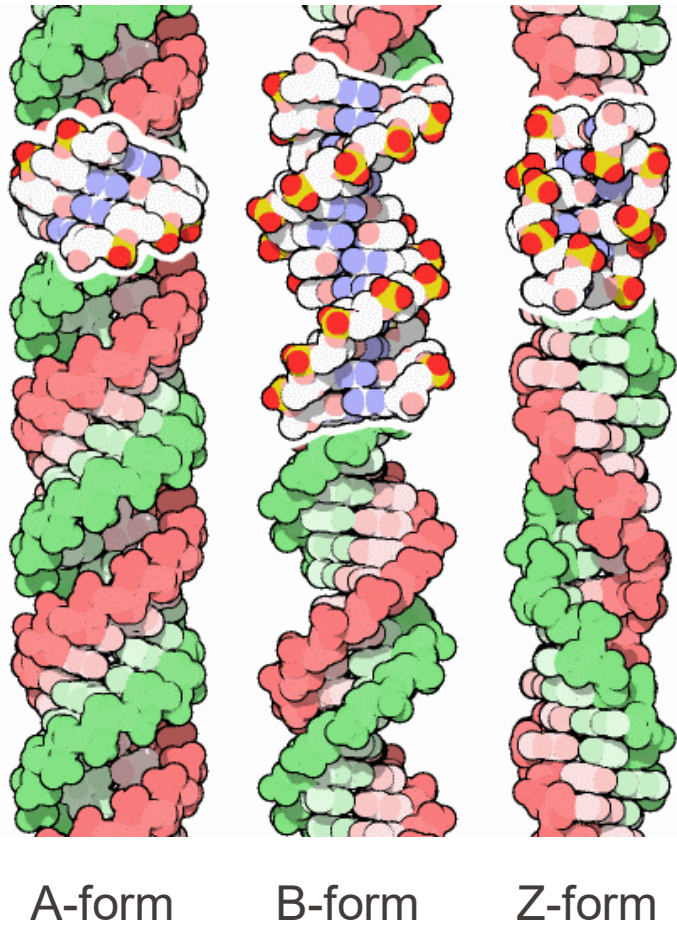
- **3' -> 5' phosphodiester linkage** allows to impose directionality to the growing strand
- By convention DNA sequences are written from **5' to the 3' end**.
- All known polymerases assemble nucleic acids in 5' -> 3' direction



Model of RNA polymerase in action
(adapted from rcsb.org).

3D assembly of DNA and RNA

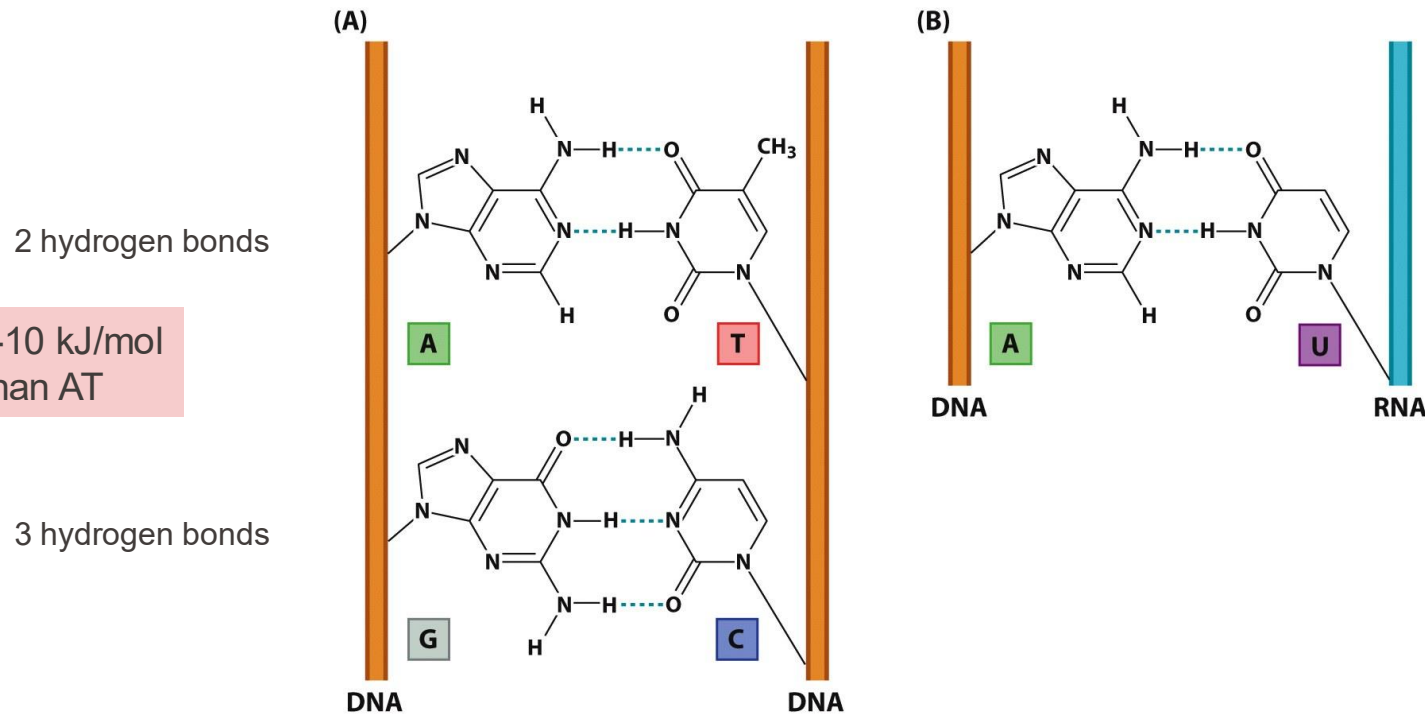
- DNA assembles into double-stranded helices
- RNA assembly varies but is usually single-stranded.



The structure of DNA

- DNA forms a **double helix** with **antiparallel strands**
- Bases face towards the inside of the helix and the phosphate backbone groups are on the outside.
- The pairing is achieved through **hydrogen bonds created between complementary bases**

GC pair is ~5-10 kJ/mol more stable than AT



The Watson-Crick base pairs: A-T, G-C and A-U

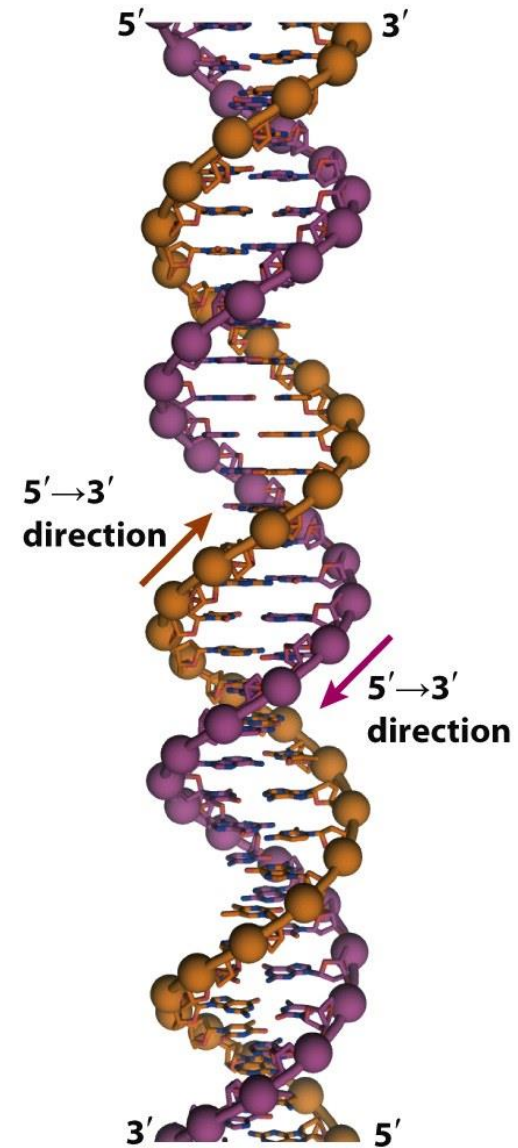


Figure 1.23 The Molecules of Life (© Garland Science 2015)

Phosphate groups
in spheres

The structure of DNA - Non-covalent Interactions

- In addition to hydrogen bonding the double helix is stabilized by **stacking of base pairs**

Beautiful example on van der Waals interactions given that the radius of **carbon** and **nitrogen** are **1.7** and **1.6 Å** respectively – the observed **rise per base-pair** is **3.3-3.4 Å**.

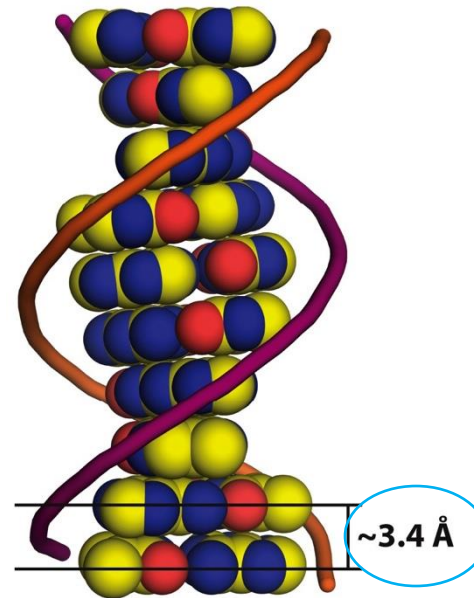
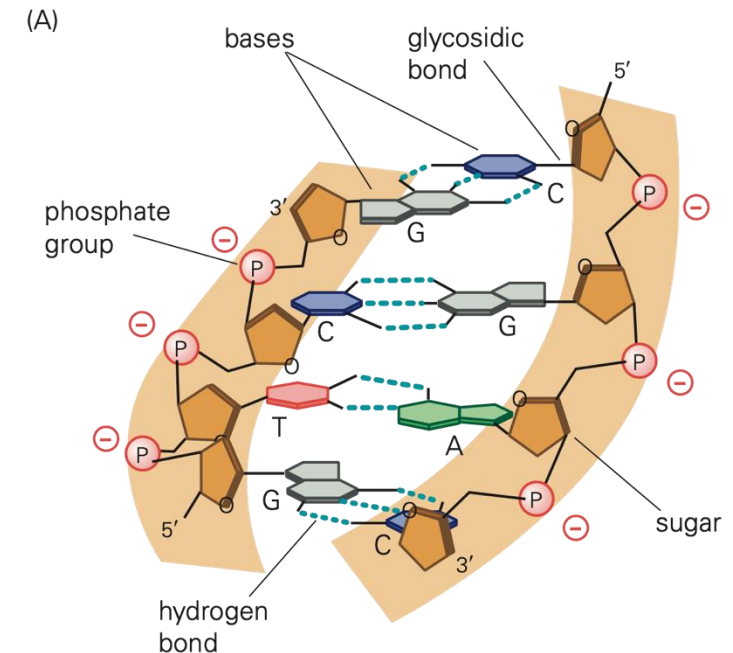


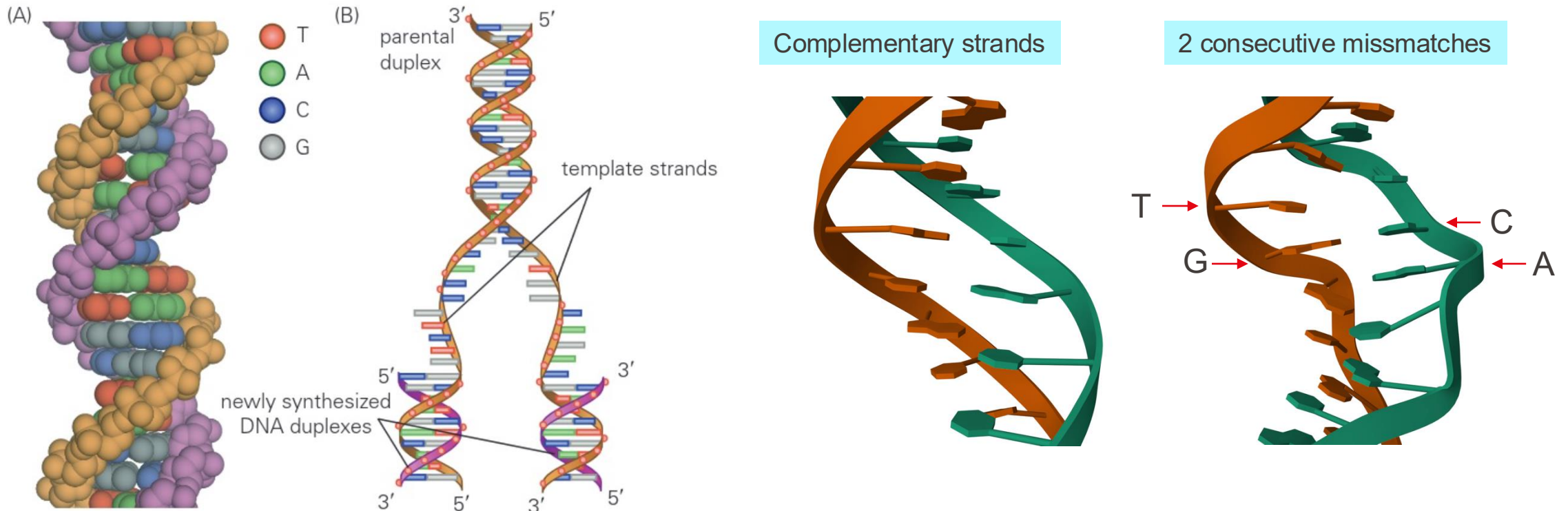
Figure 1.27 The Molecules of Life (© Garland Science 2013)



- Electrostatic interactions assure proper backbone geometry. Allowing for interactions with ions and water and minimizing repulsion between phosphates
- Base stacking is supported by solvent exclusion and **van der Waals interactions**

DNA Replication relies on base complementarity

- **DNA polymerase** is an enzyme that binds to exposed template DNA strand and adds complementary bases in 5' → 3' direction
- The end result are two DNA strands that are **reverse complements** of each other



- Mismatching base pairs result in disruption of local conformation which is detected by the DNA polymerase causing it to stall and excise the problematic base (“**proofreading**”)

Historical detour - How was DNA structure solved?

3 papers published in Nature in 1953:

- James Watson & Francis Crick
- Maurice Wilkins
- Rosalind Franklin & Raymond Gosling

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is a residue on each chain every 3.4 Å. in the z-direction. We have assumed an angle of 36° between adjacent residues in the same chain, so that the structure repeats after 10 residues on each chain, that is, after 34 Å. The distance of a ribonucleoside atom

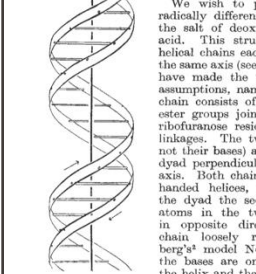
MOLECULAR STRUCTURE OF NUCLEIC ACIDS

A Structure for Deoxyribose Nucleic Acids

WE wish to suggest a structure of deoxyribose nucleic acid (DNA) which has novel features which are of biological interest.

A structure for nucleic acid has been proposed by Pauling and Corey¹. Their manuscript available to us in publication. Their model consists of two chains, with the phosphates on the outside and the bases on the inside. This structure is unsatisfactory for (1) We believe that the material in X-ray diagrams is the salt, not the free acid; the acidic hydrogen atoms it is not clear would hold the structure together, especially negatively charged phosphates near each other. (2) Some of the distances appear to be too small.

Another three-chain structure has been suggested by Fraser (in the press). In phosphates are on the outside and the inside, linked together by hydrogen bonds as described in rather ill-defined terms.



This figure is purely diagrammatic. The two ribbons symbolize the two phosphate-sugar chains, and the horizontal rods the pairs of bases holding the chains together. The vertical line marks the fibre axis.

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King's College, London. One of us (J. D. W.) has been aided by a fellowship from the National Foundation for Infantile Paralysis.

J. D. WATSON
F. H. C. CRICK

Medical Research Council Unit for the Study of the Molecular Structure of Biological Systems, Cavendish Laboratory, Cambridge, April 2.

¹Pauling, L., and Corey, R. H., *Nature*, **271**, 540 (1953); *Proc. U.S. Acad. Sci.*, **29**, 94 (1953).
²Furlberg, S., *Acta Chem. Scand.*, **2**, 534 (1952).
³Chargaff, E., for references see Sauerbrey, S., *Strawenzon*, G., and Chargaff, E., *Biochim. et Biophys. Acta*, **9**, 502 (1952).
⁴Wyatt, G. R., *J. Physiol.*, **58**, 201 (1953).
⁵Asbury, W. T., *Symp. Soc. Exp. Biol.*, **1**, Nucleic Acid, 66 (Cambridge Univ. Press, 1947).
⁶Wilkins, M. H. F., and Randall, J. T., *Biochim. et Biophys. Acta*, **10**, 192 (1953).

Molecular Structure of Deoxyribose Nucleic Acids

WHILE the biological properties of deoxyribose nucleic acid suggest a molecular structure of unusual complexity, X-ray diffraction studies described here (cf. Asbury¹) show the basic molecular configuration has great simplicity. The purpose of this communication is to describe, in a preliminary way, some of the experimental evidence for the polynucleotide chain configuration being helical, as existing in this form when in the natural state; fuller account of the work will be published shortly.

The structure of deoxyribose nucleic acid is the same in all species (although the nitrogen base radical varies considerably) in nucleoproteins, extracted cells, and in purified nucleate. The same linear group of polynucleotide chains may pack together parallel in different ways to give crystalline², semi-crystalline³ or paracrystalline material. In all cases the X-ray diffraction photograph consists of two regions, one determined largely by the regular spacing of nucleotides along the chain, and the other by the long spacings of the chain configuration. The sequence of different nitrogen bases along the chain is not visible.

Deviated paracrystalline deoxyribose nucleic acid 'structure B' in the following communication. Franklin and Gosling gives a fibre diagram as shown in Fig. 1 (cf. ref. 4). Asbury suggested that the strong 3.4-Å. reflexion corresponded to the 10 nucleotide repeat along the fibre axis. The ~34 Å. layer lines, however, are not due to a repeat of polynucleotide composition, but to the chain configuration repeat, which causes strong diffraction of the nucleotide chains have higher density than the inter-chain spaces. The absence of reflexions near the meridian immediately suggests a helical structure with axis parallel to fibre length.

Diffraction by Helices

It may be shown⁵ (also Stokes, unpublished) that the intensity distribution in the diffraction pattern of a series of points equally spaced along a helix given by the squares of Bessel functions. A uniform continuous helix gives a series of layer lines of equal intensity corresponding to the helix pitch. The intensity distribution along the nth layer line being proportional to the square of J_n , the nth order Bessel function. A straight line may be drawn approximately through

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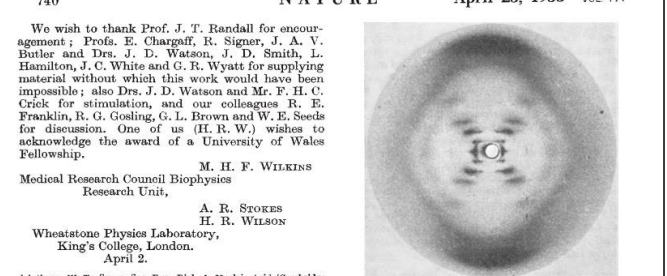
We wish to thank Prof. J. T. Randall for encouragement; Prof. E. Chargaff, R. Siger, J. A. V. Butler and Drs. J. D. Watson, J. D. Smith, L. Hamilton, J. C. White and G. R. Wyatt for supplying material without which this work would have been impossible; also Drs. J. D. Watson and Mr. F. H. C. Crick for stimulation, and our colleagues R. E. Franklin, R. G. Gosling, G. L. Brown and W. E. Seeds for discussion. One of us (H. R. W.) wishes to acknowledge the award of a University of Wales Fellowship.

M. H. F. WILKINS

Medical Research Council Biophysics Research Unit,

A. R. STOKES
H. R. WILSON

Wheatstone Physics Laboratory, King's College, London, April 2.



Sodium deoxyribose nucleate from calf thymus. Structure B

¹Asbury, W. T., *Symp. Soc. Exp. Biol.*, **1**, Nucleic Acid (Cambridge Univ. Press, 1947).
²Killey, D. P., and Oster, G., *Biochim. et Biophys. Acta*, **7**, 536 (1951).
³Wilkins, M. H. F., Gosling, R. G., and Seeds, W. R., *Nature*, **167**, 735 (1951).
⁴Asbury, W. T., and Bell, F. O., *Cold Spring Harb. Symp. Quant. Biol.*, **6**, 109 (1952).
⁵Cochran, W., Crick, F. H. C., and Vand, V., *Acta Cryst.*, **8**, 581 (1952).
⁶Wilkins, M. H. F., and Randall, J. T., *Biochim. et Biophys. Acta*, **10**, 192 (1953).

Molecular Configuration in Sodium Thymonucleate

SODIUM thymonucleate fibres give two distinct types of X-ray diagram. The first corresponds to a crystalline form, structure A, obtained at about 75 per cent relative humidity; a study of this is described in detail elsewhere¹. At higher humidities a different structure, structure B, showing a lower degree of order, appears and persists over a wide range of ambient humidity. The change from A to B is reversible. The water content of structure B fibres which undergo this reversible change may vary from 40-50 per cent to several hundred per cent of the dry weight. Moreover, some fibres never show structure A, and in these structure B can be obtained with an even lower water content.

The X-ray diagram of structure B (see photograph) shows in striking manner the features characteristic of helical structures, first worked out in this laboratory by Stokes (unpublished) and by Crick, Cochran and Vand². Stokes and Wilkins were the first to propose such structures for nucleic acid as a result of direct studies of nucleic acid fibres, although a helical structure had been previously suggested by Furlberg (thesis, London, 1949) on the basis of X-ray studies of nucleosides and nucleotides.

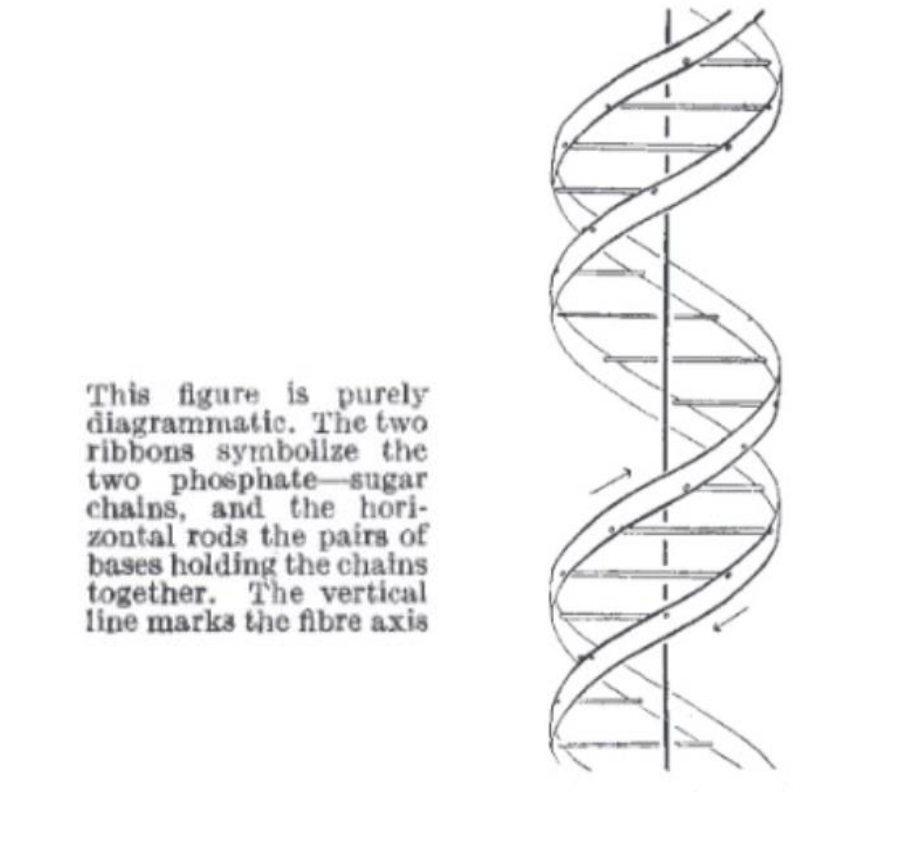
While the X-ray evidence cannot, at present, be taken as direct proof that the structure is helical, other considerations discussed below make the existence of a helical structure highly probable. Structure B is derived from the crystalline structure A when the sodium thymonucleate fibres take up quantities of water in excess of about 40 per cent of their weight. The change is accompanied by an increase of about 30 per cent in the length of the fibre, and by a substantial re-arrangement of the molecule. It therefore seems reasonable to suppose that in structure B the structural units of sodium thymonucleate (molecules or groups of molecules) are relatively free from the influence of neighbouring

$$I_n = J_n^2(2\pi r r) \exp i n(\psi + \frac{1}{2}\pi),$$

where $J_n(u)$ is the nth-order Bessel function of u , r is the radius of the helix, and R and ψ are the radial and azimuthal co-ordinates in reciprocal space; this expression leads to an approximately linear array of intensity maxima of the type observed, corresponding to the first maxima in the functions J_1, J_2, J_3 , etc.

If, instead of a smooth helix, we consider a series of residues equally spaced along the helix, the transform in the general case treated by Crick, Cochran and Vand is more complicated. But if there is a whole number, m , of residues per turn, the form of the transform is as for a smooth helix with the addition, only, of the same pattern repeated with its origin at heights $m\tau, 2m\tau, \dots$ etc. (τ is the fibre-axis period).

In the present case the fibre-axis period is 34 Å, and the very strong reflexion at 3.4 Å. lies on the tenth layer line. Moreover, lines of maxima radiating from the 3.4-Å. reflexion as from the origin are visible on the fifth and lower layer lines, having a J_m maximum coincident with that of the origin series on the fifth layer line. (The strong outer streaks which apparently radiate from the 3.4-Å. maximum are not, however, so easily explained.) This suggests strongly that there are exactly 10 residues per turn of the helix. If this is so, then from a measurement of R_n , the position of the first maximum on the nth layer line (for $n \leq 5$), the radius of the helix, can be obtained. In the present instance, measurements of R_1, R_2, R_3 and R_4 all lead to values of r of about 10 Å.



This figure is purely diagrammatic. The two ribbons symbolize the two phosphate-sugar chains, and the horizontal rods the pairs of bases holding the chains together. The vertical line marks the fibre axis.

Historical detour - What was known at the time?

- Entering 1950s there was a pretty good understanding of the chemical nature of nucleotides and resulting DNA, including all relevant covalent bonds between building blocks.
- Additionally, Erwin Chargaff in 1950 published his findings on the ratio of nucleotides, that ultimately led to the establishment of Chargaff rules.

• Chargaff rules:

$$\%A \approx \%T$$

$$\%G \approx \%C$$

$$\%(A+T) \neq \%(G+C)$$

University of Bern



Rudolf Signer

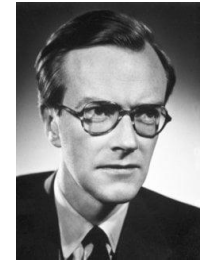
Kings College London



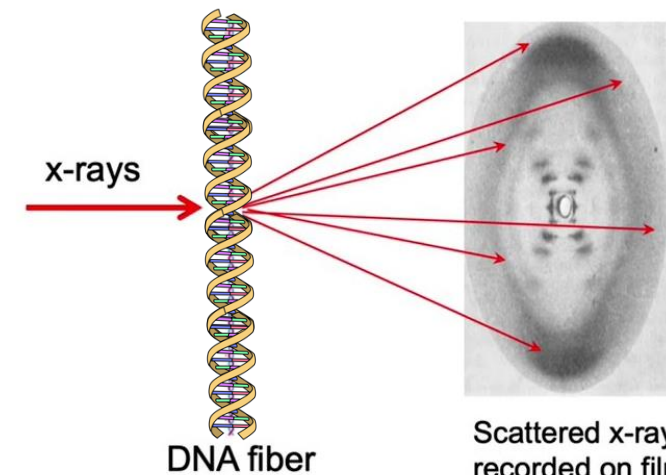
Rosalind Franklin



Raymond Gosling



Maurice Wilkins



Isolation of calf DNA

DNA fiber

Scattered x-rays recorded on film

Historical detour - The Secret of Image 51

- Spots in this diffraction image represent a plane in DNA molecule that regularly repeats leading to constructive interference
- The X and Y axis have the units of 1/distance and can be connected to the DNA planes by Bragg's law

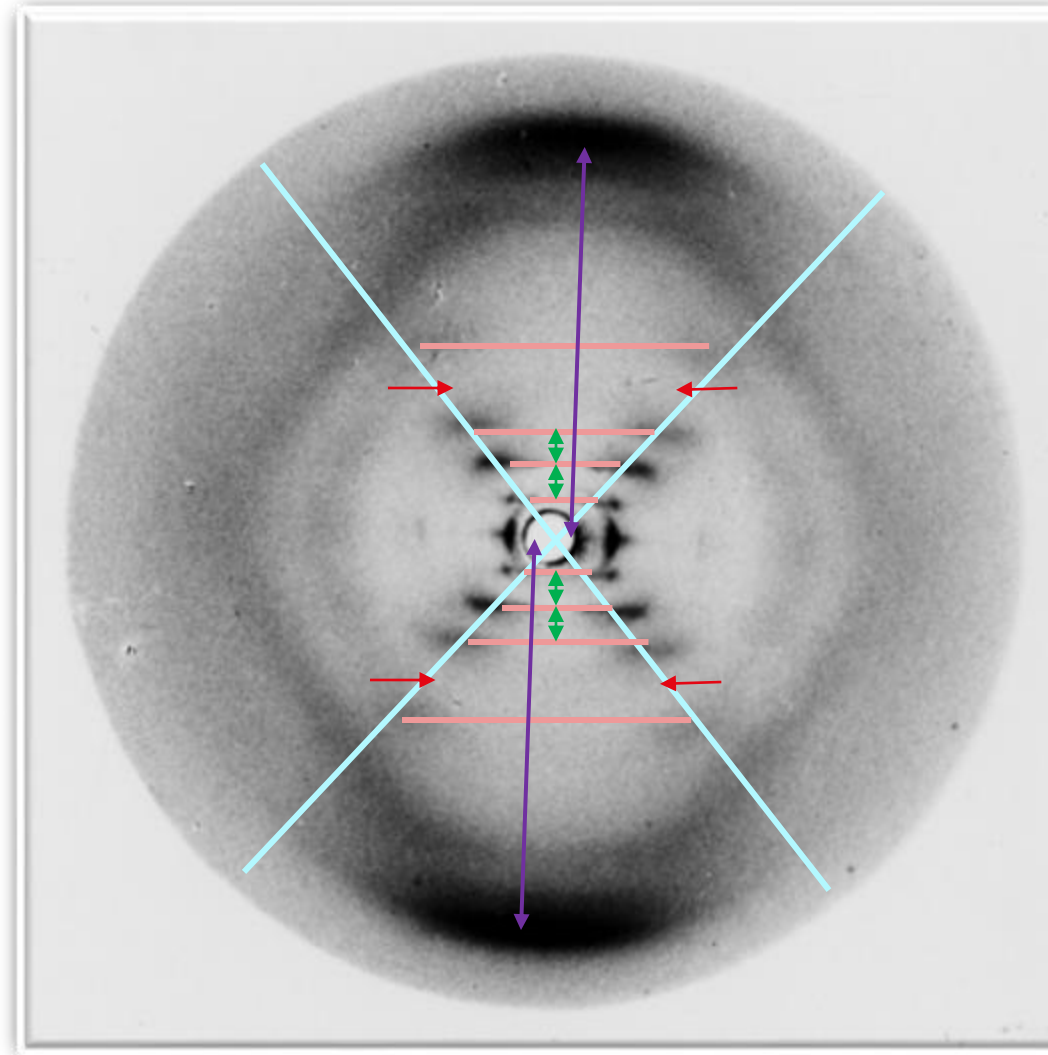
• The X-shape indicates a helix - known based on symmetry operators for helical assemblies

• The horizontal distances can be used to calculate helix width - Resulting in **$\sim 20\text{\AA}$**

• The spacing of vertical spots corresponds to one rise of the helix ("helical pitch") - Resulting in **$\sim 34\text{\AA}$**

• The position of the large peak corresponds to distance between large chemical groups (i.e., bases) - Resulting in **$\sim 3.4\text{\AA}$**

• Missing 4th layer indicates absence of perfect regularity expected for single helix - Must be more than one strand (but even number)



Rosalind Franklin's X-ray image of DNA ("Photo 51")

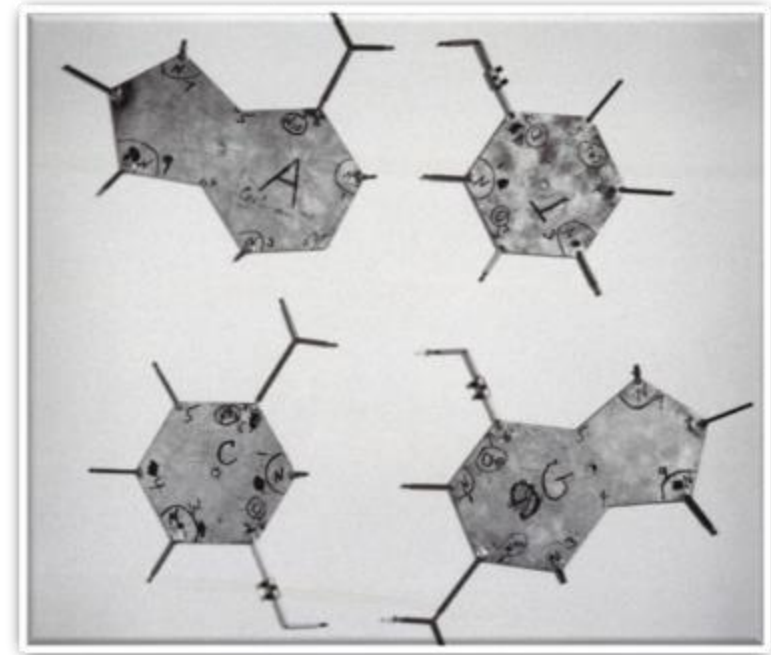
Historical detour - Crick and Watson

- James Watson and Francis Crick were junior scientists (PhD student and a postdoc) specializing in molecular modeling at Cambridge, interested in the DNA structure problem
- Controversially, Franklin decided to leave the King's College and her data was shared to Watson and Crick without her permission



DNA model designed by Crick and Watson

Crick and Watson integrated Chargaff's rules into model building assuming that the bases must pair in certain way:



Historical detour - James Watson and base pairing



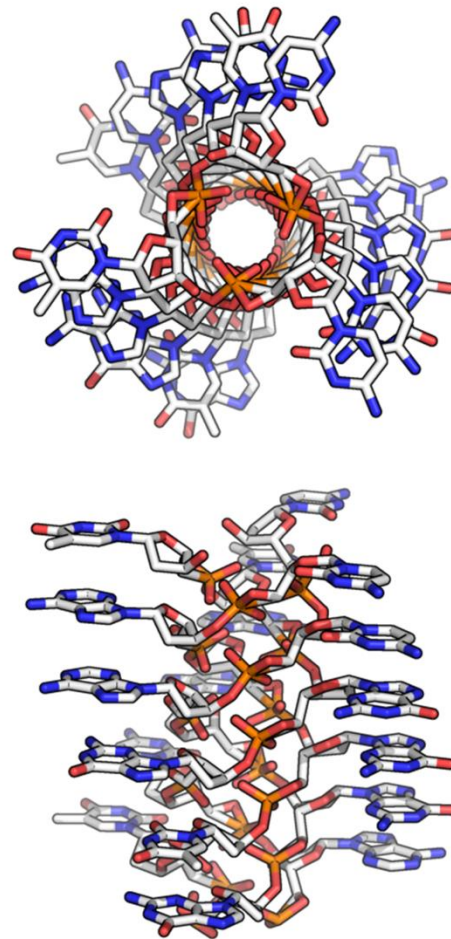
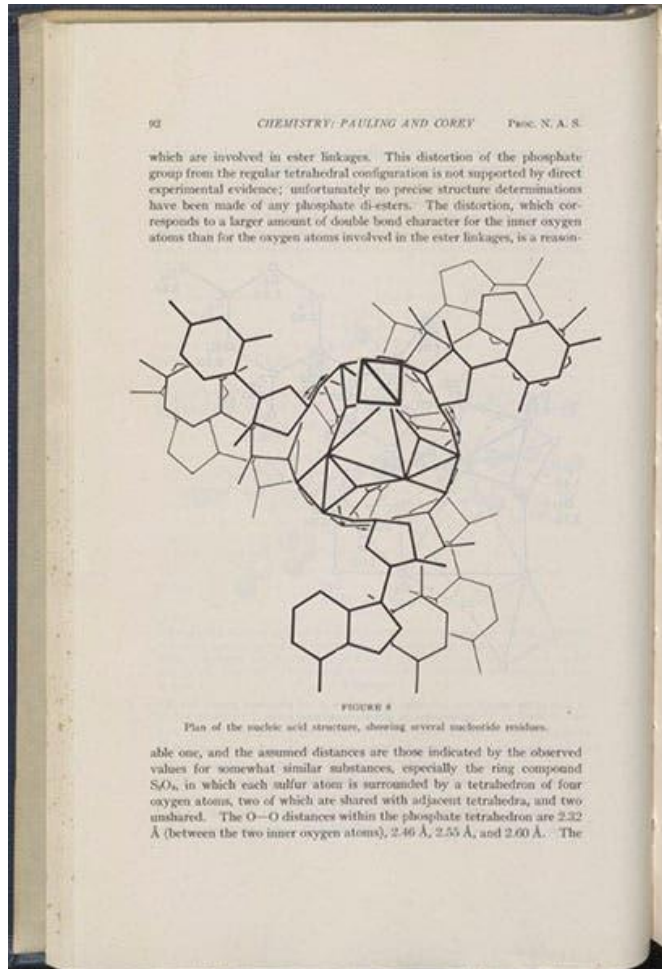
Historical detour - Nobel Prize in 1962

- James Watson, Francis Crick and Maurice Wilkins received the Nobel Prize in Physiology and Medicine in 1962
- Interestingly, Max Perutz and John Kendrew, who were the mentors of Watson and Crick, also won a Nobel Prize in the same year (Chemistry)
- Rosalind Franklin passed away in 1958 and her contribution was not acknowledged until several decades later



Historical detour - Linus Pauling model

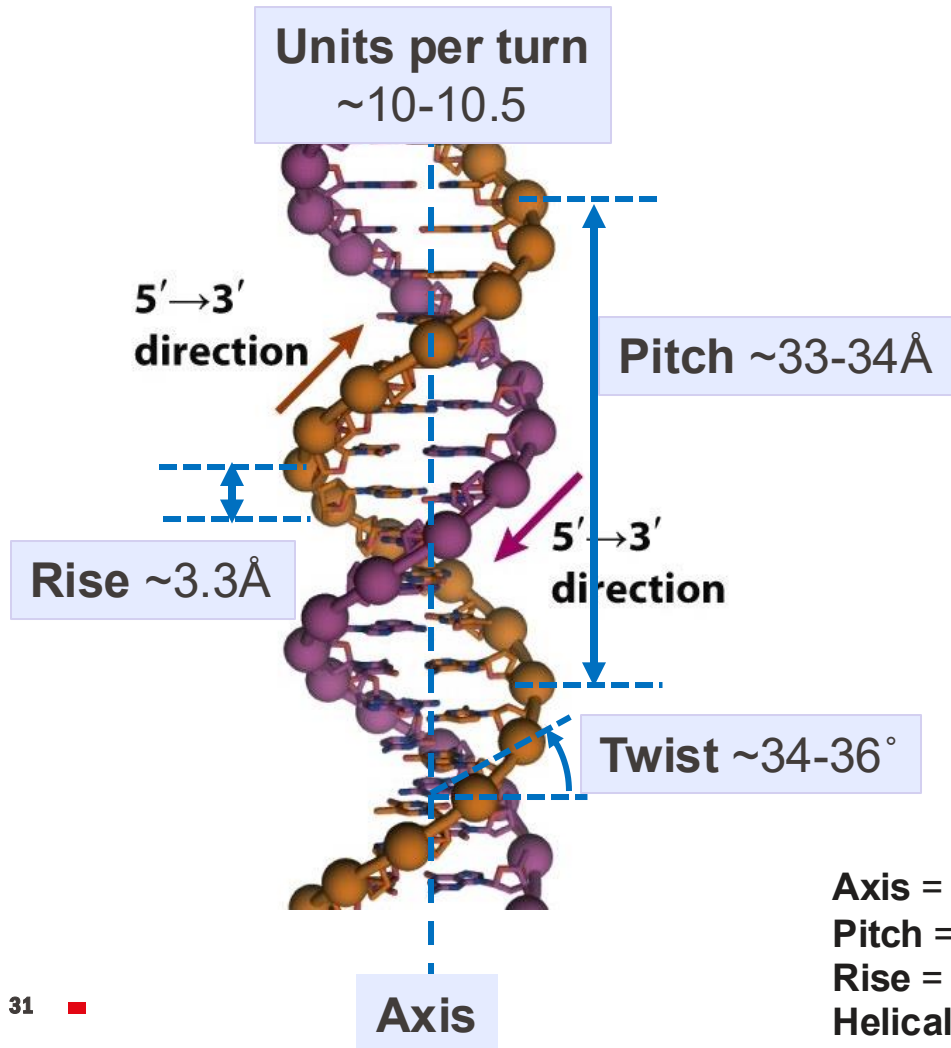
- Relying on incomplete X-ray data, Linus Pauling (2-times Nobel Laureate) proposed a structure of DNA as a triple helix



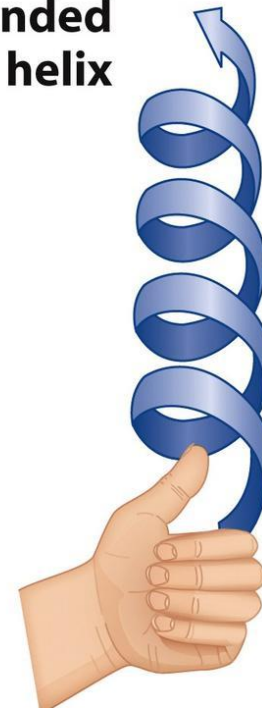
- The Pauling model proposed that all phosphate groups are in the center, while the bases face outwards
- But there was no proposed solution for resolving the negative charge build-up in the middle
- By his own admission, this proposed structure was his “greatest scientific misstep”

The structure of DNA - Helix geometry

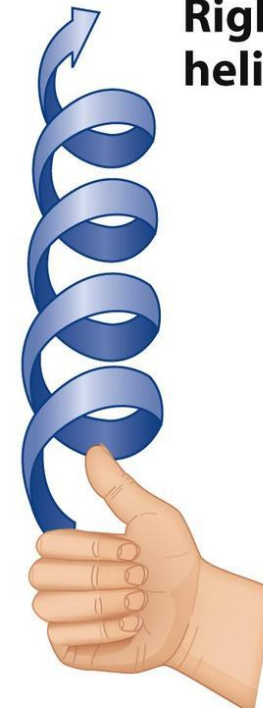
- DNA helix is a repeating unit defined by a **helical axis**, **pitch**, **twist**, and **rise**
- Two strands together wind up to form a **right-handed** double-helix



Left-handed
helix



Right-handed
helix



Axis = helical axis is an imaginary line around which the two strands wind.

Pitch = The distance along the helical axis required for one complete turn of the DNA double helix.

Rise = The distance the helix extends along its axis when you go from one base pair to the next.

Helical twist = The angle of rotation between successive base pairs around the helix axis.

Different DNA assembly forms



Rosalind Franklin

- DNA can assemble into 3 major forms: **A, B and Z**
- Their geometries are very different (e.g., **Z-helix is left-handed**)

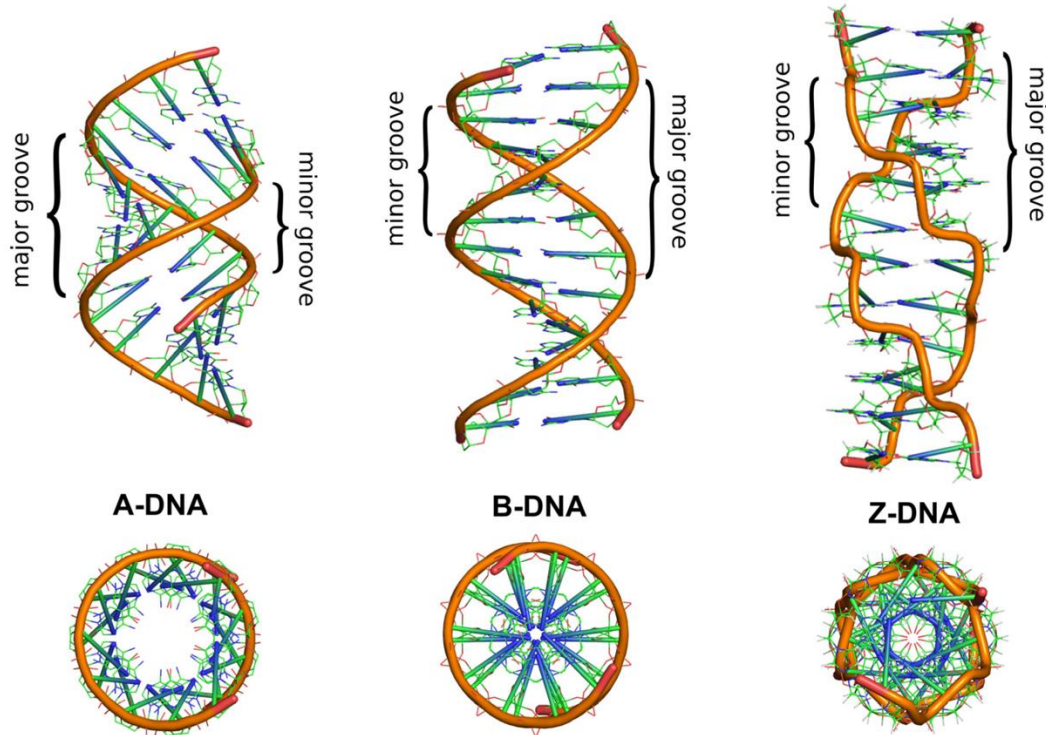


Table 2.1 Structural features of A-, B-, and Z-form helices.

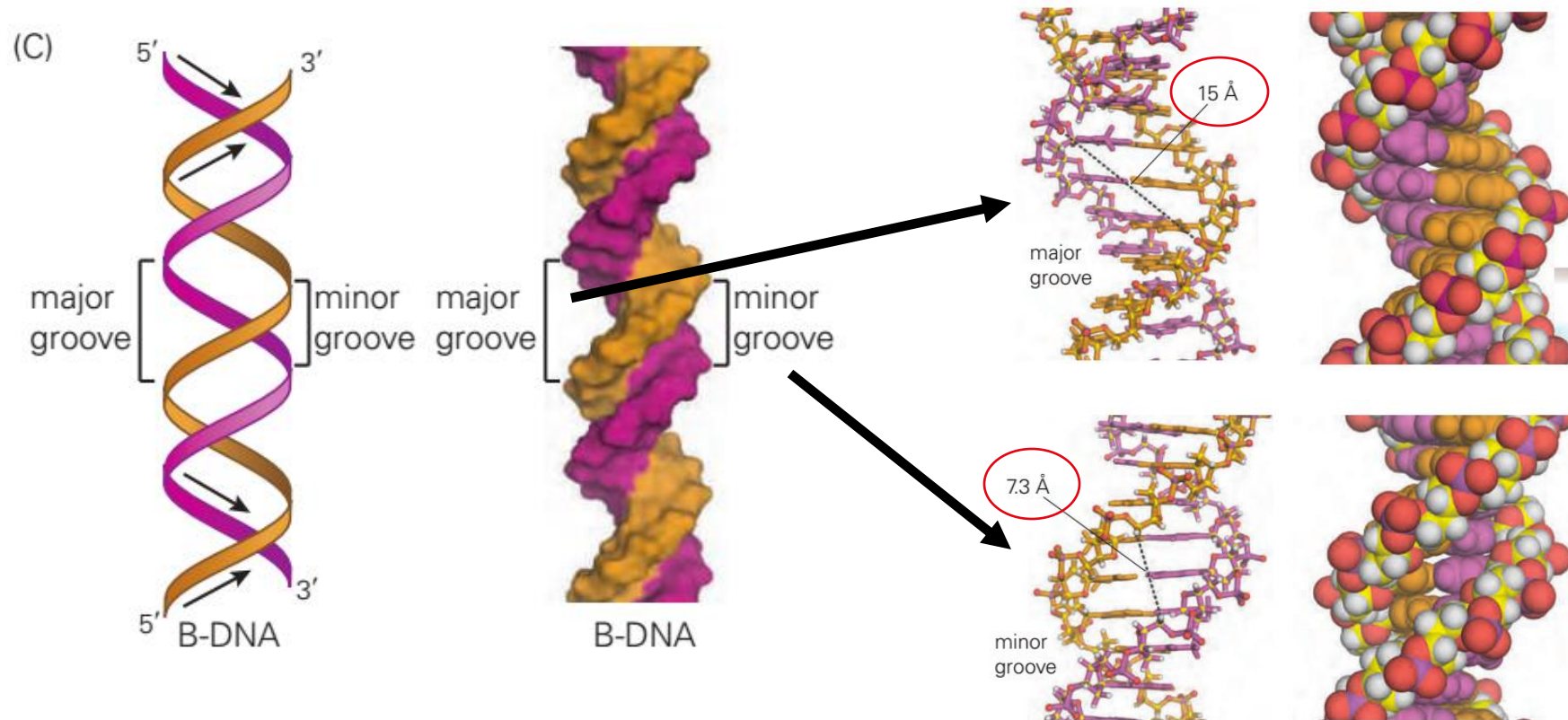
| Helical form | A | B | Z |
|---|--|---|---|
| Helical sense | Right | Right | Left |
| Diameter | ~ 26 Å | ~ 20 Å | ~ 18 Å |
| Base pairs per turn | ~ 11 | ~ 10 | ~ 12 |
| Helical twist (rotation per base pair for A and B, per two-base repeat for Z) | ~ 34° | ~ 36° | ~ 60° (CpGp) |
| Helix pitch (rise per helical turn) | ~ 25 Å | ~ 33 Å | ~ 46 Å |
| Helix rise (along helix axis; per base pair for A and B, per two-base repeat for Z) | ~ 2.3 Å | ~ 3.3 Å | ~ 7.4 Å (CpGp) |
| Base tilt (with respect to helix axis) | ~ 20° | ~ 0° | ~ - 9° |
| Base orientation (with respect to sugar) | Anti | Anti | C anti/G syn |
| Base pair positions (helix axis indicated by black dot) | | | |
| Features of base pair positions | Base pairs displaced from axis; deep major groove, less accessible | Base pairs on axis; both major and minor grooves accessible | Base pairs stick out into the major groove, the minor groove is deep and narrow |

(Adapted from R.E. Dickerson et al., and M.L. Kopka, *Science* 216: 475–482, 1982. With permission from AAAS.)

- **The major form in solution is B**, while A and Z are less prevalent (and require special conditions)
- Relative locations of C2' and C3' carbons in deoxyribose are responsible for A and B forms

Major and minor grooves on DNA

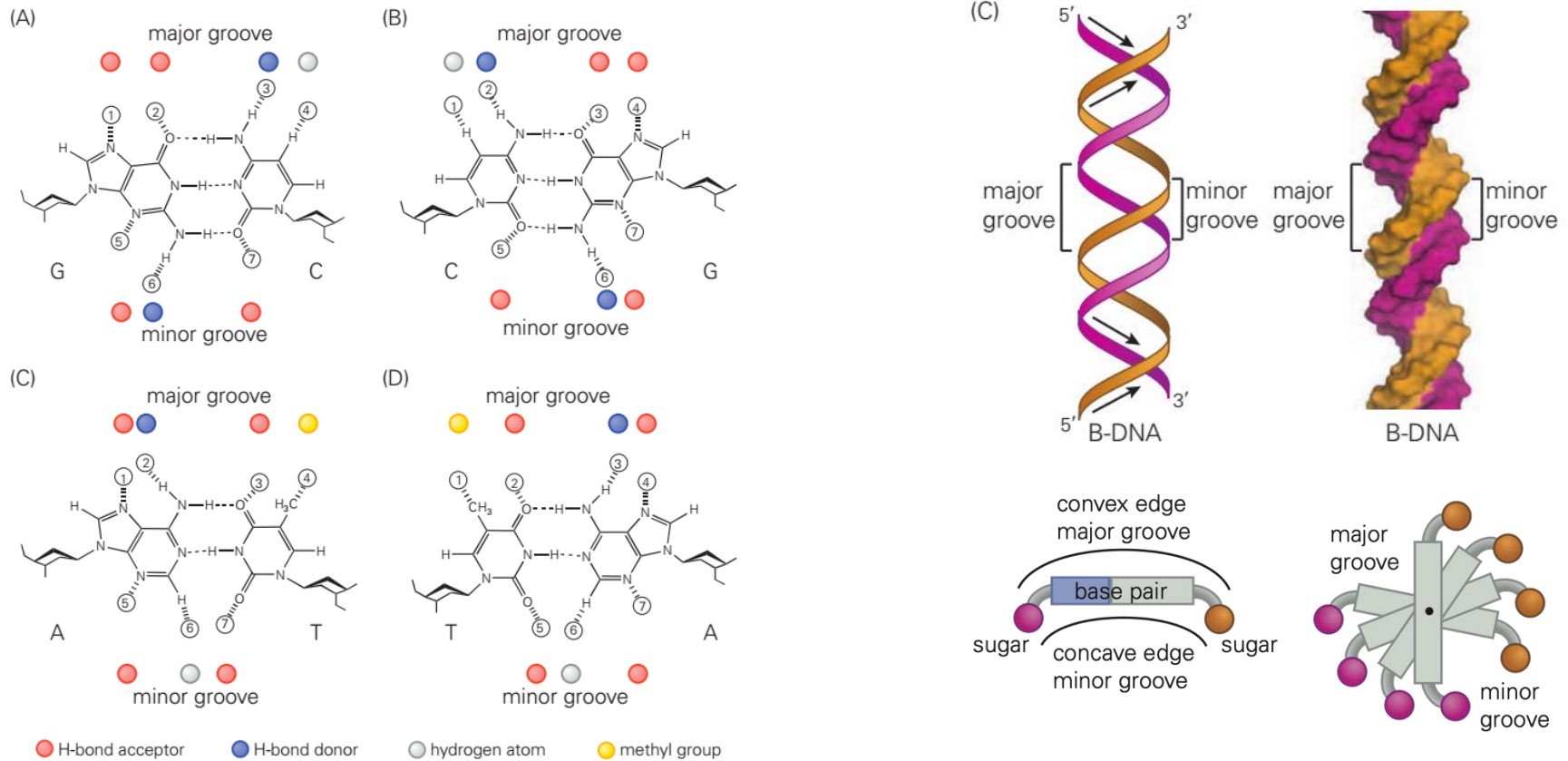
- In addition to sequence, one of the most important features of DNA double helices are the **grooves**.



- The **major** and **minor** groove present very distinct structural features
- Very important for the **recognition of DNA by proteins**.

Major and minor grooves on DNA

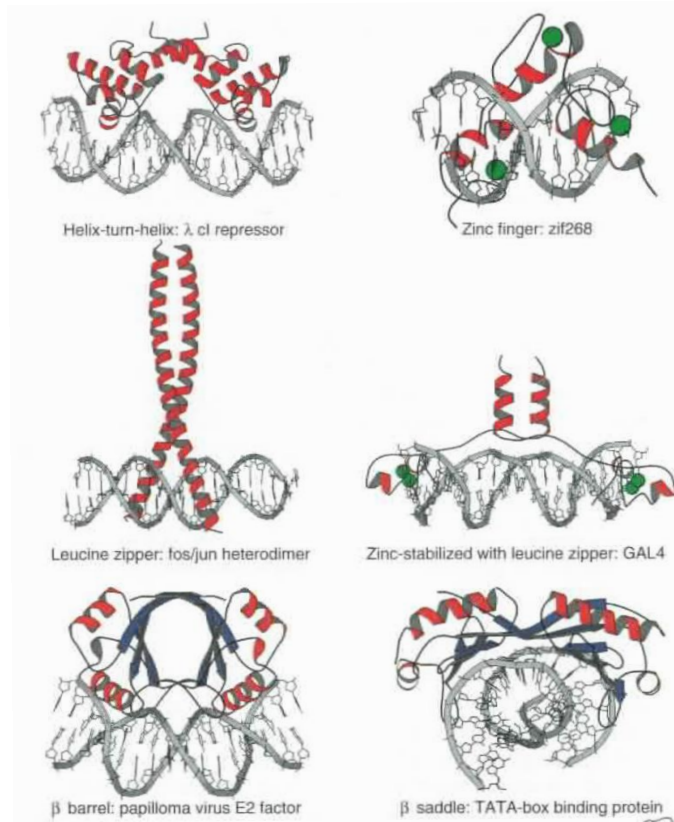
- Potential **secondary interaction sites** at the edges of Watson-crick base pairs



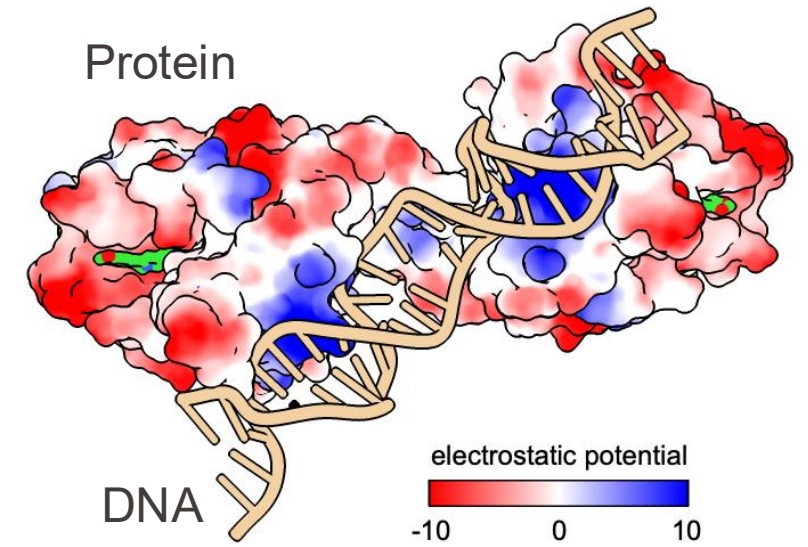
- The major and minor grooves can be identified by looking at the connections of the base pairs with the sugars.
- Major groove on the **convex edge** and the **minor groove** in on the concave edge.

DNA-binding proteins

- DNA-binding proteins primarily target the major groove (Mgr) or a combination of major and minor (Mm).
- Understanding how networks of DNA binding proteins impact cell processes is an essential component of immunology, developmental, cancer and systems biology.



| Complex | Binding Motif ^a | Binding Groove ^b | Details of Complex |
|------------------------------|----------------------------|-----------------------------|---|
| λ repressor | HTH | Mgr | Canonical HTH; homodimers; 2 helices of Cro dimer cradle Mgr, stabilized by direct H-bond and vdW contacts; little DNA distortion. About 90° bend. |
| CAP repressor | HTH | Mgr | Indirect, water-mediated base contacts. |
| try repressor | HTH | Mgr | α-helices inserted in mgr. |
| Purine rep. | HTH | Mm | Homocox domains bind as monomers. |
| Yeast MATα2 | HTH | Mgr | |
| Zif268 | Zn | Mgr | Zinc finger subfamily; each Zn finger recognizes 3 bps. |
| GATA-1 | Zn | Mm | Transcription factors subfamily; single domain coordinated by 4 cysteines. |
| GAL4 | Zn | Mgr | Metal binding subfamily; each of two Zn ions, coordinated by 6 cysteines, recognizes 3 bps. |
| GCN4 | Leu/Zip | Mgr | Canonical; basic region/leucine zipper (α helices) motif; slight DNA bending. |
| fos/jun | Leu/Zip | Mgr | α-helices resemble GCN4; unstructured basic region folds upon DNA binding. |
| fos/jun/NEAT | Leu/Zip | Mgr | α-helices bend to interact with NEAT. |
| MutI | β-ribbon | Mgr | Two anti-parallel β-strands in Mgr; bends each DNA end by 25°. |
| papillomavirus E2 DNA target | β-barrel | Mgr | Dimered β-sheets form an 8-strand β-barrel dimer interface with 2 α-helices in Mgr; strong tailored fit for every base of the recognition element; best DNA; compressed mgr; DNA target crystallized without protein. |
| TBP | β-saddle | mgr | Tm-β-strand saddle binds in Mgr; significant distortion, ~90° bend. |
| ρ/β tumor supp. | Loop/other | Mm | Binds to DNA via protruding loop and helix anchored to anti-parallel β-barrel. |
| SRY | Loop/other | mgr | Intoxinc intercalated into mgr. |
| NEAT | Loop/other | Mm | Flexible binding loop stabilized by DNA. |
| bisoxanes | Loop/other | Mm | Nonspecific PO ₄ interactions. |
| distamycin (drug) | | mgr | Selective to AT bps; binds in mgr without distortion. |



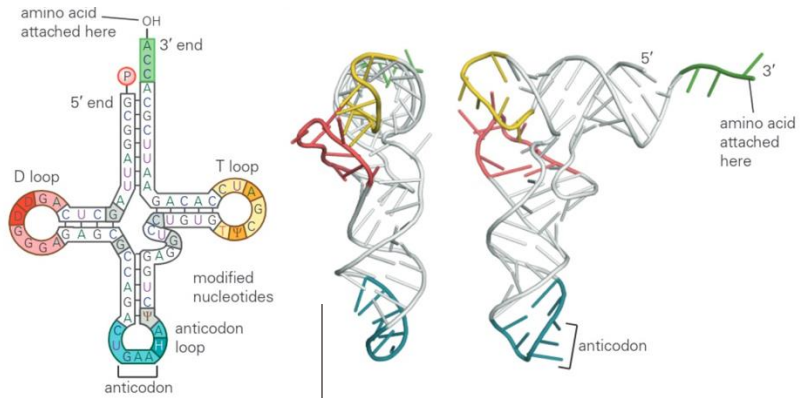
DNA-binding proteins typically engage the DNA backbone using positively charged amino acids



Differences in assembly of DNA and RNA

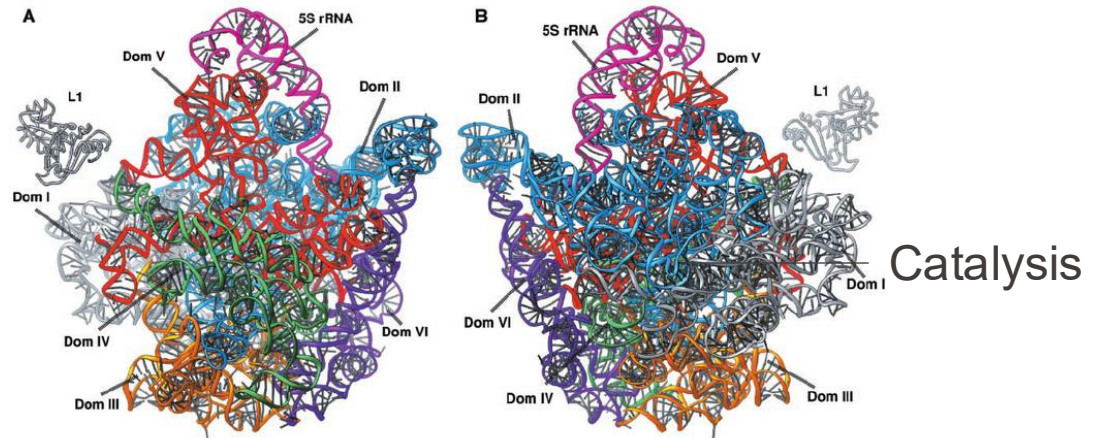
- Due to the unique chemical restraints of the (i) ribose sugar and the (ii) single-stranded nature in most cases, RNA can take on many diverse forms and, consequently, has more biological functions compared to DNA

transfer (t) RNA

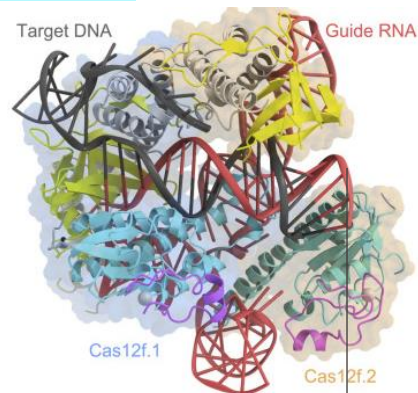


Amino-acid carrier

Ribosomal RNA



Guide RNA



Defending

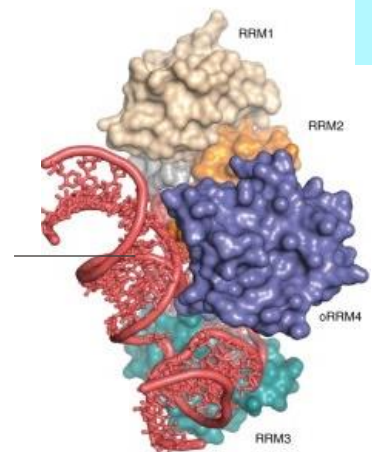
Riboswitch



Ligand sensing

snRNA

Scaffolding
(and catalysis)

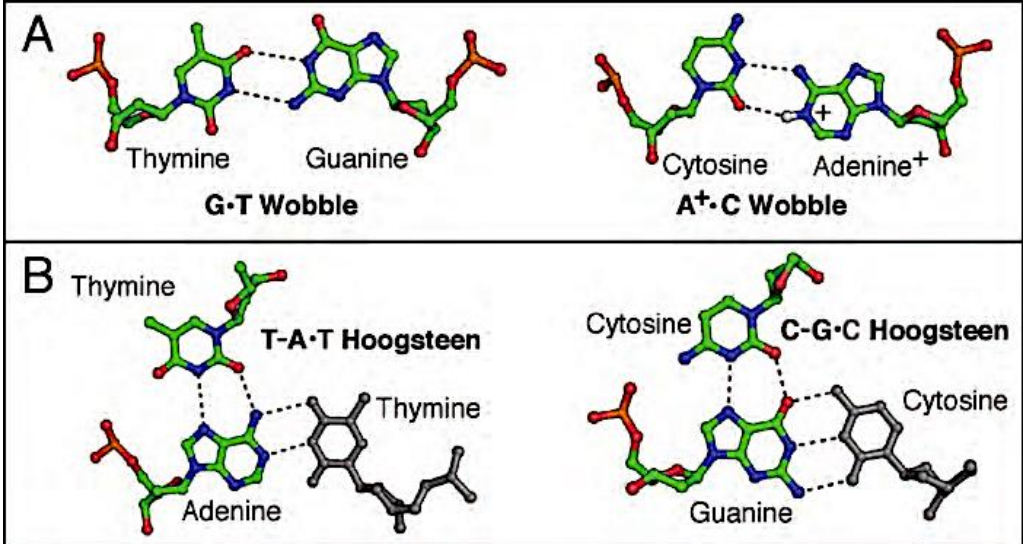
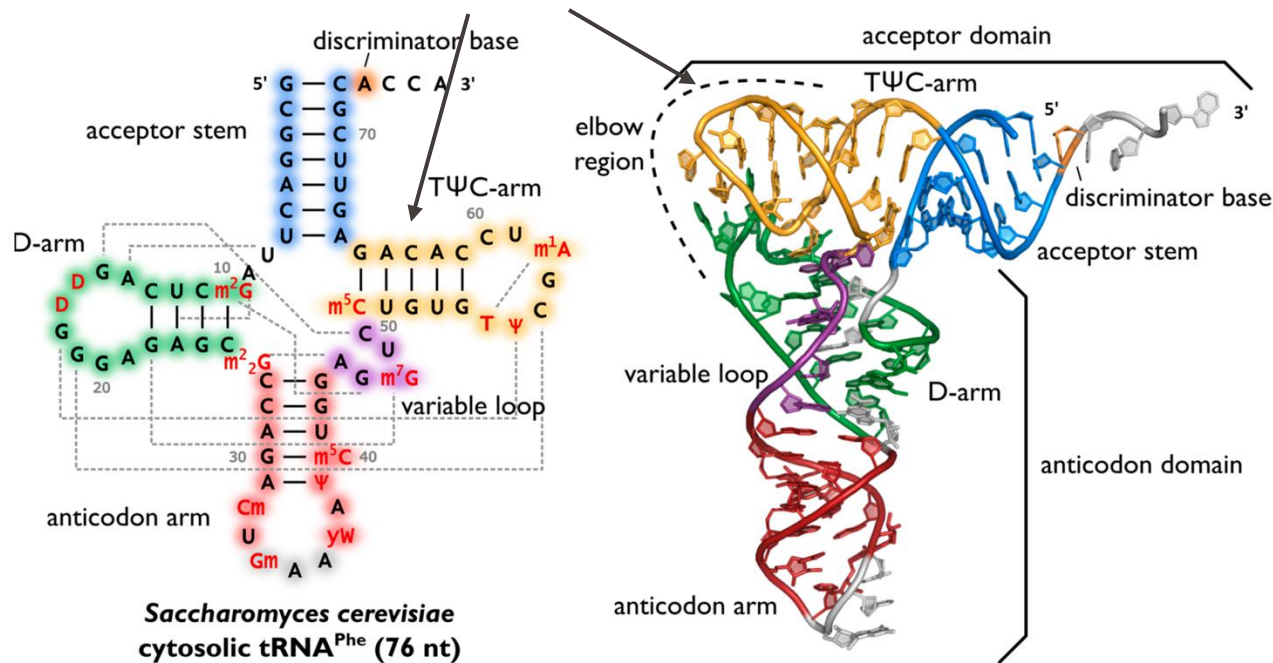


RNA can also form partial double-helices

- Partial complementary regions in **RNA** drive the assembly of **shorter double helices** which ultimately dictate the 3D assembly of the entire RNA molecule and its function.
- The conformation is further stabilized by **non-canonical base pairing** interactions. These can also occur in DNA but are more common in RNA.

Complementary regions assemble into short double-stranded helices

Non-Watson-Crick base pairing improves stability of non-complementary regions

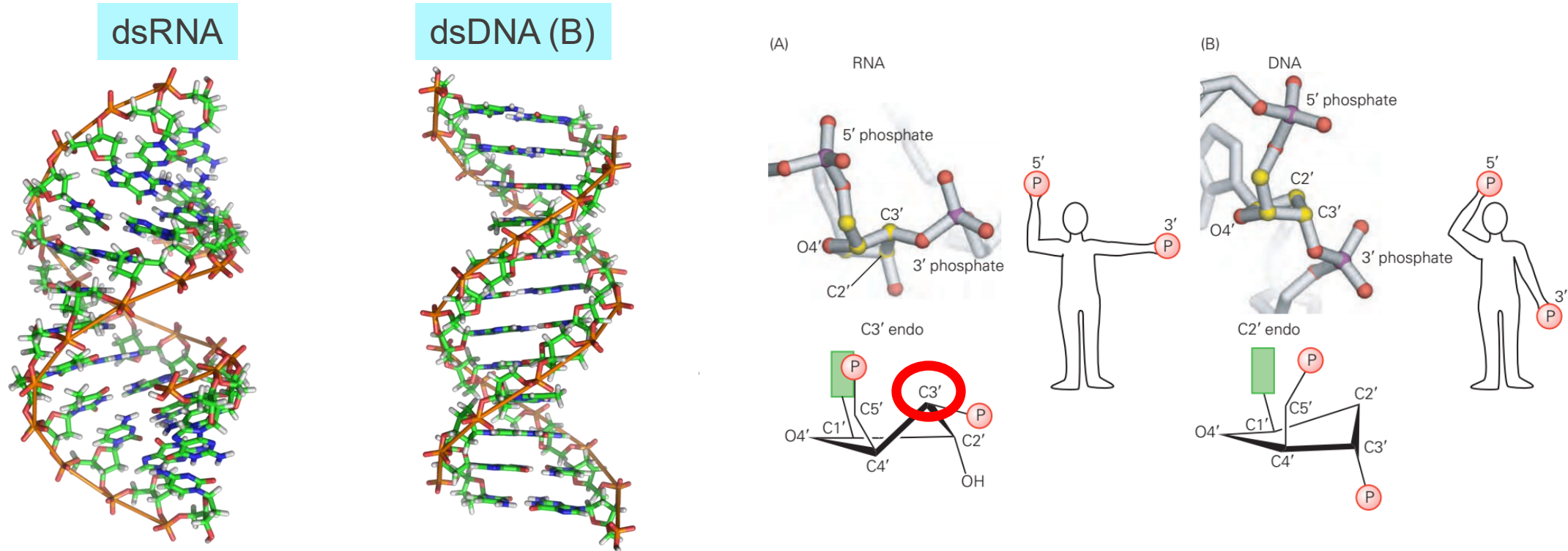


A few examples, but many more exist...

No need to remember these for exam. You just need to be aware of their existence.

Differences in assembly of dsDNA and dsRNA

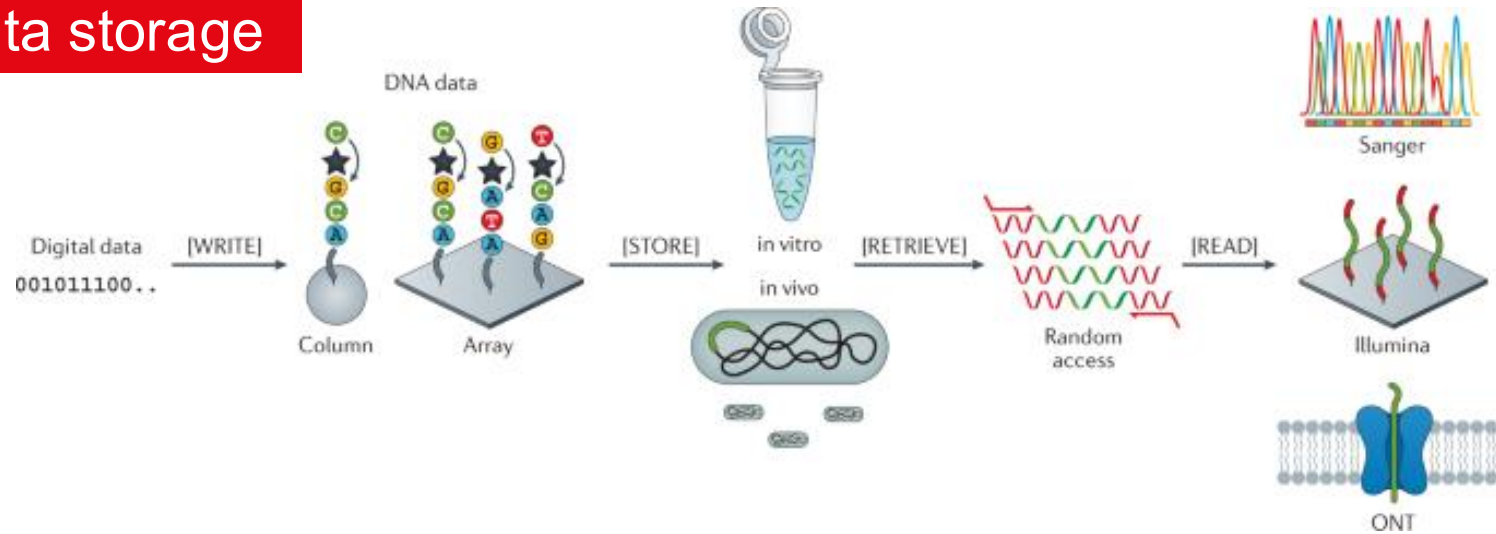
- Double-stranded helices of RNA can be formed locally in molecules like tRNA, but they can also be found in some viruses (e.g., rotavirus) where dsRNA represents the viral genome



- For RNA the hydroxyl group influences the preferred conformational state of the sugar (**C3'-up state**) which impacts how it packs into a helix. In that sense it is similar to the **A-form** of DNA

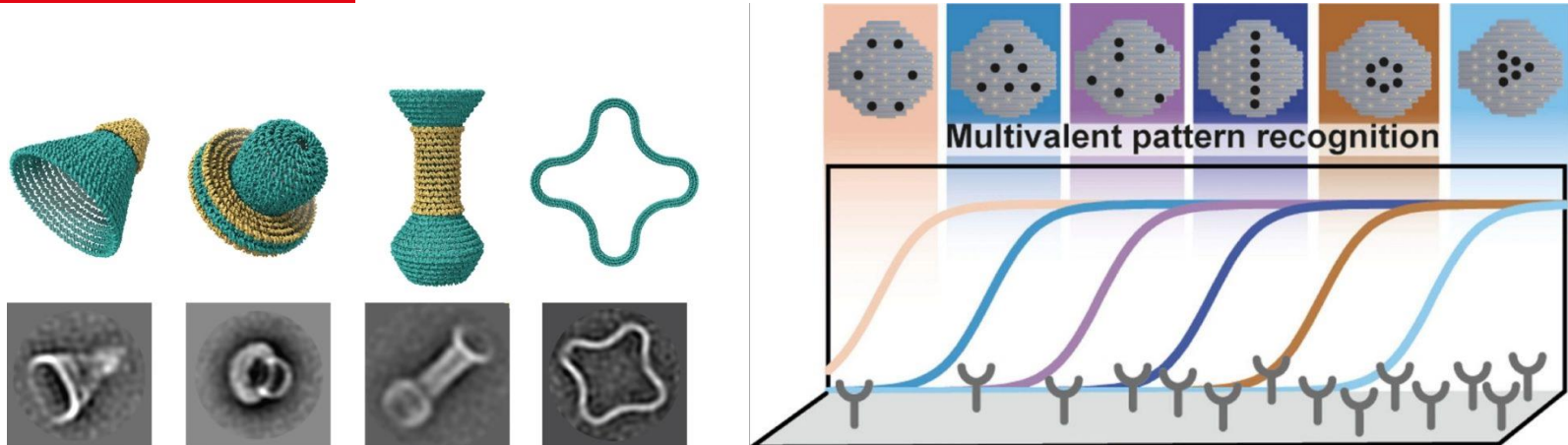
DNA in biotechnology

Data storage



- Ceze, L., Nivala, J. & Strauss, K. Molecular digital data storage using DNA. *Nat Rev Genet* **20**, 456–466 (2019). <https://doi.org/10.1038/s41576-019-0125-3>

DNA origami

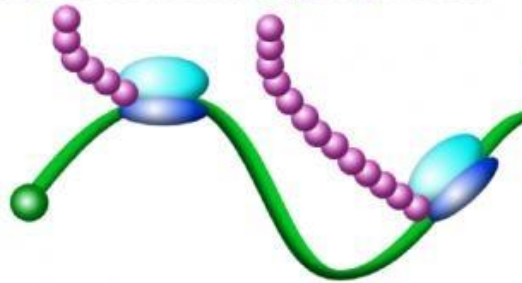


- Hale Bila, Kaltrina Paloja, Vincenzo Caroprese, Artem Kononenko, Maartje M.C. Bastings, *American Chemical Society*, (2022)
- <https://pubs.acs.org/doi/10.1021/jacs.2c08529#>

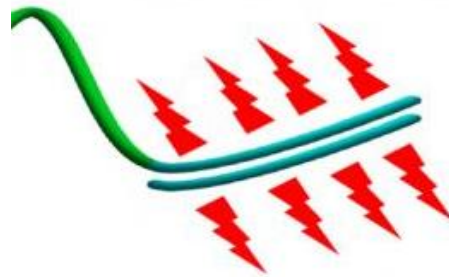
RNA in biotechnology

Vaccines and adjuvants

Expression of antigen protein

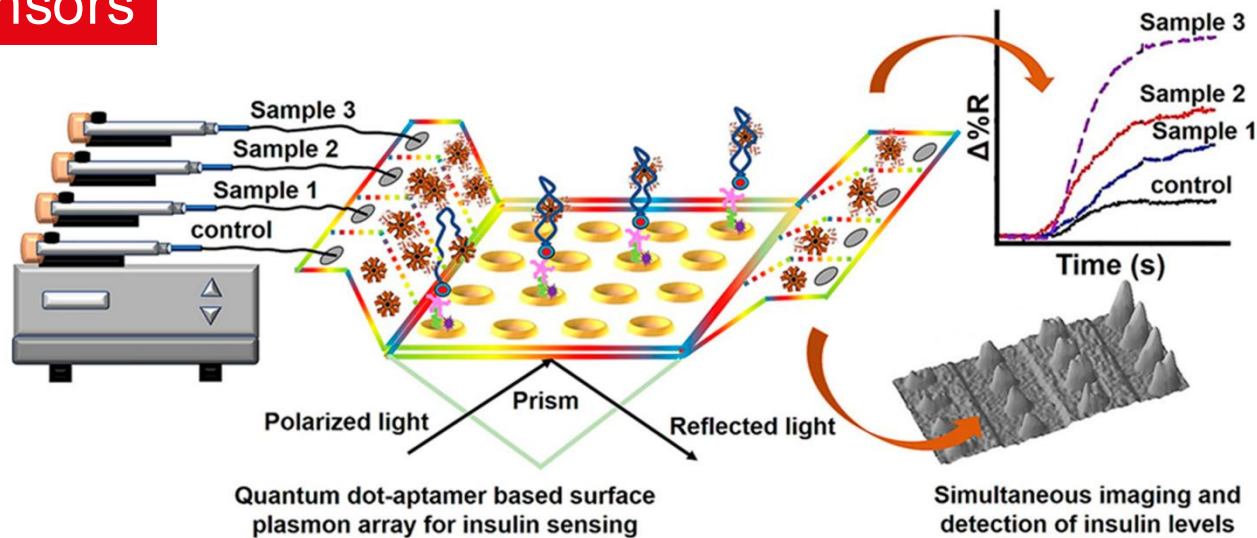


Immuno-stimulation by double stranded RNA structure



- Abhijeet Girish Lokras, Thomas Rønnemoes Bobak, Saahil Sandeep Baghel, Federica Sebastiani, Camilla Foged, *Advanced Drug Delivery Reviews*, Vol 213, 2024.
- <https://www.sciencedirect.com/science/article/pii/S0169409X24002412>

Biosensors



- Nako Nakatsuka, Kelly J. Heard, Alix Faillétaz, Dmitry Momotenko, János Vörös, Fred H. Gage & Krishna C. Vadodaria, *Molecular Psychiatry* volume 26, pages2753–2763, 2021.
- <https://www.nature.com/articles/s41380-021-01066-5>

Nucleic acids – Take Home messages

- **DNA and RNA are the informational polymers** in the cell – encode genetic information in a way that can be read by macromolecular machines, to direct the synthesis of other molecules.
- Nucleotides have **pentose sugars** attached to **nitrogenous bases** and **phosphate** groups.
- The nucleotide bases in DNA and RNA are substituted **pyrimidines** or **purines**.
- **4 deoxyribonucleotides** in DNA (A,T,G,C) and **4 ribonucleotides** in RNA (A,U,C,G)
- **DNA and RNA are synthesized in 5' to 3' direction** by sequential reactions that are driven by hydrolysis of nucleotide triphosphates
- DNA forms a **double helix** with antiparallel strands (**direct** and **reverse complement** strands)
- Double helix involves complementary base pairing (A-T and C-G) and is stabilized by, **hydrogen bonds, base pair stacking and electrostatic interactions**
- B-form DNA allows sequence specific **recognition of the major groove by proteins**. Each base pair has a unique set of interacting elements in the major groove but not in the minor groove.
- RNA is typically single-stranded, but it can also assemble into shorter or longer helical structures.

Next week's lecture
Introduction to Lipids



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